

Nucleobase-containing compounds evoke behavioural, olfactory, and transcriptional responses in model fishes

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Abstract

The sensory system of animals detects a massive and unknown array of chemical cues that evoke a diversity of physiological and behavioural responses. One group of nitrogen-containing carbon ring chemicals—nucleobases—are thought to be involved in numerous behaviours yet have received little attention. We took a top-down approach to examine responses evoked by nucleobases at behavioural, tissue, and gene expression levels. Fish generally avoided nucleobases, and this behaviour, when observed, was driven by purines but not pyrimidines. At the tissue level, olfactory neuron generator potential responses tended to be concentration specific and robust at concentrations lower than amino acid detection ranges. In terms of gene expression, more than 2000 genes were significantly upregulated following nucleobase exposure, some of which were expected (e.g., genes involved in purine binding) and some of which were not (e.g., tubulin-related genes). Humanized RNA pathway analysis showed that we had exposed the animal to a nucleobase. Our data indicate that responses to nucleobase-containing compounds may be highly structure based and are evident from changes in behaviour to mRNA expression. Many of these responses were surprising, and all provide numerous routes for further research endeavour.

Key words: nucleotides, nucleosides, ATP, olfaction, RNA sequencing, zebrafish, goldfish

Introduction

A huge diversity of chemicals can evoke a vast array of sensory responses. Animals use chemical stimuli ranging in complexity from the simple to the elaborate to facilitate activities from feeding to mating. One of the more intriguing classes of understudied chemical stimuli are nucleobase-containing compounds, i.e., compounds having carbon rings with nitrogen (**Table 1**). Among these compounds are the sugar-containing nucleosides, which include deoxyribonucleic acids (DNA), and the phosphorylated nucleotides, which include adenosine triphosphate (ATP). Nucleotides are released by bacteria ([Okabayashi et al. 1963](#); [Demain et al. 1965](#); [Saier et al. 1975](#)) and animals; the latter may excrete these compounds naturally ([Murad and Pak 1972](#); [Sato et al. 1981](#))

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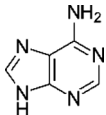
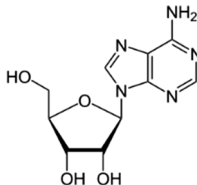
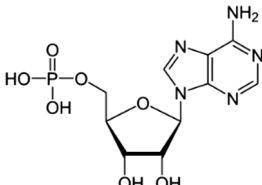
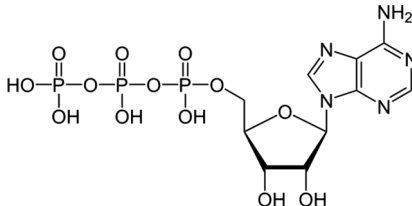
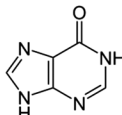
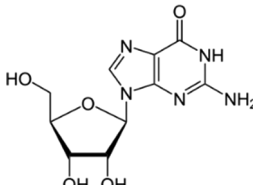
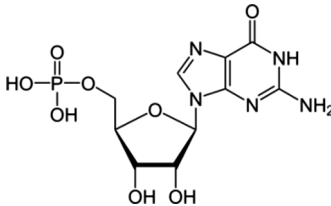
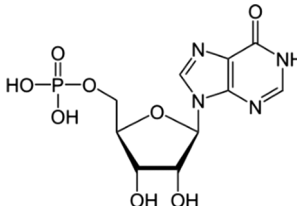
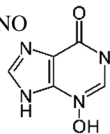
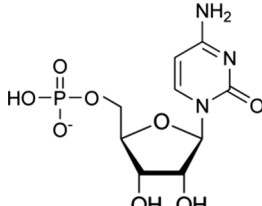
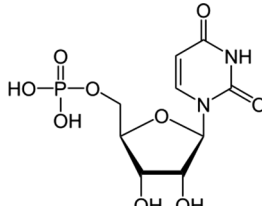
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Table 1. Purine and pyrimidine compounds used in this study.

Compounds				
Purines				
Adenine	Adenosine	AMP	ATP	
				
Hypoxanthine	Guanosine	GMP	IMP	
				
H3NO				
				
Pyrimidines				
		CMP	UMP	
				

Note: Images from [Wikimedia Commons](#). AMP, adenosine-5'-monophosphate monohydrate; ATP, adenosine-5'-triphosphate disodium salt hydrate; GMP, guanosine-5'-monophosphate disodium salt hydrate; CMP, cytidine-5'-monophosphate disodium salt; UMP, uridine-5'-monophosphate disodium salt; IMP, inosine-5'-monophosphate disodium salt hydrate.

or involuntarily through injury (Brown et al. 2000, 2001). Nucleobases may serve as sensory stimuli that evoke a range of responses in animals.

In general, nucleobases evoke “eat or be eaten” behaviours. In several hematophagous insects, including the bed bug (*Cimex lectularius* Linnaeus, 1758) and the kissing bug (*Rhodnius prolixus* Stål, 1859), nucleotides are potent initiators of gorging behaviour (Friend and Smith 1971; Smith and Griend 1976; Romero and Schal 2014). In abalone (*Haliotis discus* Reeve, 1846) and rainbow trout (*Oncorhynchus mykiss* (Walbaum, 1792)), complex nucleotide mixtures evoked feeding (Mearns et al. 1987; Miyasaki and Harada 2003). A single nucleotide (inosine monophosphate) evoked feeding behaviour in various fishes (Carr et al. 1996). In contrast, a single nucleobase (hypoxanthine 3-*N*-oxide; H3NO) evoked the opposite of feeding—an alarm response—in several fishes (Brown et al. 2000, 2001, 2003; Parra et al. 2009; Mathuru et al. 2012).

In bed bugs and kissing bugs, reducing adenosine phosphorylation diminished responses (Friend and Smith 1971; Smith and Griend 1976; Romero and Schal 2014). These findings were supported in the Caribbean spiny lobster (*Panulirus argus* (Latreille, 1804)) (Carr et al. 1986). Presumably, the greater the number of phosphate groups, the “fresher” and more stimulatory the stimulus, as ATP will hydrolyze in water. For zebrafish (*Danio rerio* (Hamilton, 1822)), un-phosphorylated adenosine evoked a variety of changes in swimming activity (Shamchuk and Tierney 2012). There may also be differences among species; for example, adenine-containing compounds induced feeding in adult rainbow trout but not Atlantic salmon parr (*Salmo salar* Linnaeus, 1758) (Mearns et al. 1987). Additionally, the alarm response evoked by H3NO in ostariophysan fishes appeared to be absent in a salmonid (Brown et al. 2003). Of interest is understanding how specific nucleobase structures evoke behaviours.

This study explored the behavioural and physiological responses of larval zebrafish to nucleobases. We expected that some nucleobases would evoke attraction or feeding (appetitive) responses, whereas others would evoke avoidance or alarm. Appetitive responses may be apparent as increased foraging (Jones and Hara 1985; Mearns et al. 1987), which may be evident as increased swimming activity (Jones and Hara 1985; Lindsay and Vogt 2004; Braubach et al. 2009; Bhinder and Tierney 2012). Avoidance responses are self-evident and well described in fishes (Tierney 2016); alarm responses may be apparent as erratic and rapid movements and periods of “freezing” behaviour (Brown et al. 2000, 2001; Speedie and Gerlai 2008; Parra et al. 2009; Mathuru et al. 2012). We examined behaviour in embryonic zebrafish (5–7 d post-fertilization; dpf) in part because they are a model vertebrate, and so their responses may have relevance to both basic and applied studies, and in part to explore developmental changes in responses. The olfactory epithelium of embryonic zebrafish begins to function as early as 3 dpf (Lindsay and Vogt 2004; Li et al. 2005; Vitebsky et al. 2005). Although the olfactory system is functional, it is undergoing change. Odourant receptor expression changes over time (Barth et al. 1996; Argo et al. 2003), and the activity of the olfactory bulb increases with age (Argo et al. 2003; Li et al. 2005). Furthermore, not all receptors have the same onset of expression, nor do they increase at the same rate (Barth et al. 1996; Argo et al. 2003). Calcium imaging of olfactory bulb activity indicated odourant-induced activity was greater after just 2 d of development (5 vs. 3 dpf) (Li et al. 2005). Additionally, as embryonic fish develop, they have increased movement ability and need for food (Kimmel et al. 1995; Colwill and Creton 2011). All of this developmental change suggests that behavioural responses to odourants may vary.

The three objectives of this study were to determine (1) whether nucleobases evoked behaviours in a structure-dependent manner, (2) whether behaviours varied over embryonic development, and (3) whether nucleobases evoked physiological responses in the sensory epithelium or at the transcriptional level.

Materials and methods

Animals

Adult AB genotype zebrafish were housed in a stand-alone system (Aquaneering, USA) at the University of Alberta. Water was passed through coarse filters, fluidized filter beads, and carbon cartridges and underwent UV sterilization. Fish were maintained at 28 °C on a light:dark cycle of 14 h of light and 10 h of darkness per day (LD 14:10). Adult fish were fed twice daily with a supplemented trout chow mixture including blood worms (Omega Sea, China), spirulina flakes (Cobalt Aquatics, USA), and TetraMin flakes (Tetra, Germany). Zebrafish breeding pairs were separated by dividers until the morning breeding window to reduce temporal variation in development (Westerfield 1993). Embryos collected from breeding pairs were reared in embryo medium (EM; details in the [Supplementary Material](#)) at 28.5 °C under a LD 14:10 cycle until they reached the desired testing age (5–7 dpf) (Westerfield 1993). The EM was refreshed daily and dead or morphologically abnormal

embryos were removed. Adult goldfish (*Carassius auratus auratus* (Linnaeus, 1758)) with a length of 10–15 cm ($N = 7$) were obtained from Mt. Parnell Fisheries (Mercersburg, Pennsylvania) and held in dechlorinated municipal tap water at 18 °C in flow-through aquaria at the University of Alberta Aquatics Facility. Goldfish were held for >3 weeks prior to experiments. Fish held in the aquatics facility were subjected to a LD 16:8 cycle and were fed daily at 0900. Animal use was approved by the University of Alberta's Animal Care and Use Committee (AUP #052).

Chemical stimuli

Avoidance–attraction was tested with adenine, adenosine, adenosine-5'-monophosphate monohydrate (AMP), adenosine-5'-triphosphate disodium salt hydrate (ATP), guanosine, guanosine-5'-monophosphate disodium salt hydrate (GMP), cytidine-5'-monophosphate disodium salt (CMP), uridine-5'-monophosphate disodium salt (UMP), inosine-5'-monophosphate disodium salt hydrate (Sigma, Ontario), and H3NO, which was synthesized (details in the [Supplementary Material](#)). Stock solutions of nucleobase compounds were prepared at 1×10^{-3} mol/L in EM, adjusted to pH 7.20 ± 0.05 , and stored at -20 °C prior to testing. A nucleotide mixture was also tested, which consisted of UMP, AMP, CMP, and GMP combined at equal molarity for a final stock concentration of 4×10^{-3} mol/L in EM. For experiments, aliquots were thawed and solutions were prepared in EM prior to testing at final concentrations of 1×10^{-5} and 4×10^{-5} mol/L for individual compounds and the mixture, respectively. Freezing of odourant stock solutions was previously shown not to reduce their efficacy ([Parra et al. 2009](#)). For H3NO, a concentration of 5×10^{-9} mol/L was used because it was shown to be the most effective concentration in adult zebrafish ([Parra et al. 2009](#)).

Behavioural responses

There are at least two methods to examine changes in swimming behaviour evoked by odourants: introduce an odourant into a static environment and allow the fish to find it, or introduce an odourant into a flow-through environment where a fish has no choice but to find it. Each method has pros and cons. The advantage of the first method is that it allows a fish to choose to move towards or away from the stimulus (i.e., make a choice), but the disadvantage is that the introduced water may itself be a stimulus ([Shamchuk and Tierney 2012](#)). With the second method, fish are forced to experience an odourant (i.e., at the expense of choice). We used both methods but report primarily on the avoidance data because behavioural changes were largely absent in the flow-through apparatus.

Avoidance–attraction responses

Nucleobase avoidance or attraction was tested in embryonic zebrafish of 5, 6, and 7 dpf. An avoidance–attraction trough was used in which fish were acclimated to a central area before being given access to side chambers via removable inserts (all three chambers were of equal size; see [Fig. S1](#)). The trough was contained within a curtained enclosure to limit disturbances from external stimuli. For experiments, 50 mL of EM was added to the trough and the barriers were inserted. Experimental fish ($n = 9$ –11 per trial; all larvae were tested only once) in 7 mL of EM were transferred by pipette to the middle chamber of the apparatus. The water level across the three chambers was equalized during the 20-min acclimation and the test solutions were pipetted into adjacent chambers and mixed with the pipette tip (the stimulus side was randomized). Post-acclimation, barriers were removed, which caused little water disturbance, and overhead video was recorded for 10 min (SX-920C-HR camera; Matco, Quebec, Canada). Eight to 10 trials were carried out for each stimulus. This apparatus was first validated using embryos and food extracts and manual scoring of embryo movement ([Fig. 1](#)). All behavioural trials were carried out between the hours of 0800 and 1800, ensuring light exposure only within the rearing light cycle. Zebrafish larvae may exhibit greater baseline movement in the morning versus the afternoon ([MacPhail et al. 2009](#)), and so stimulus testing time was randomized to prevent temporal biases.

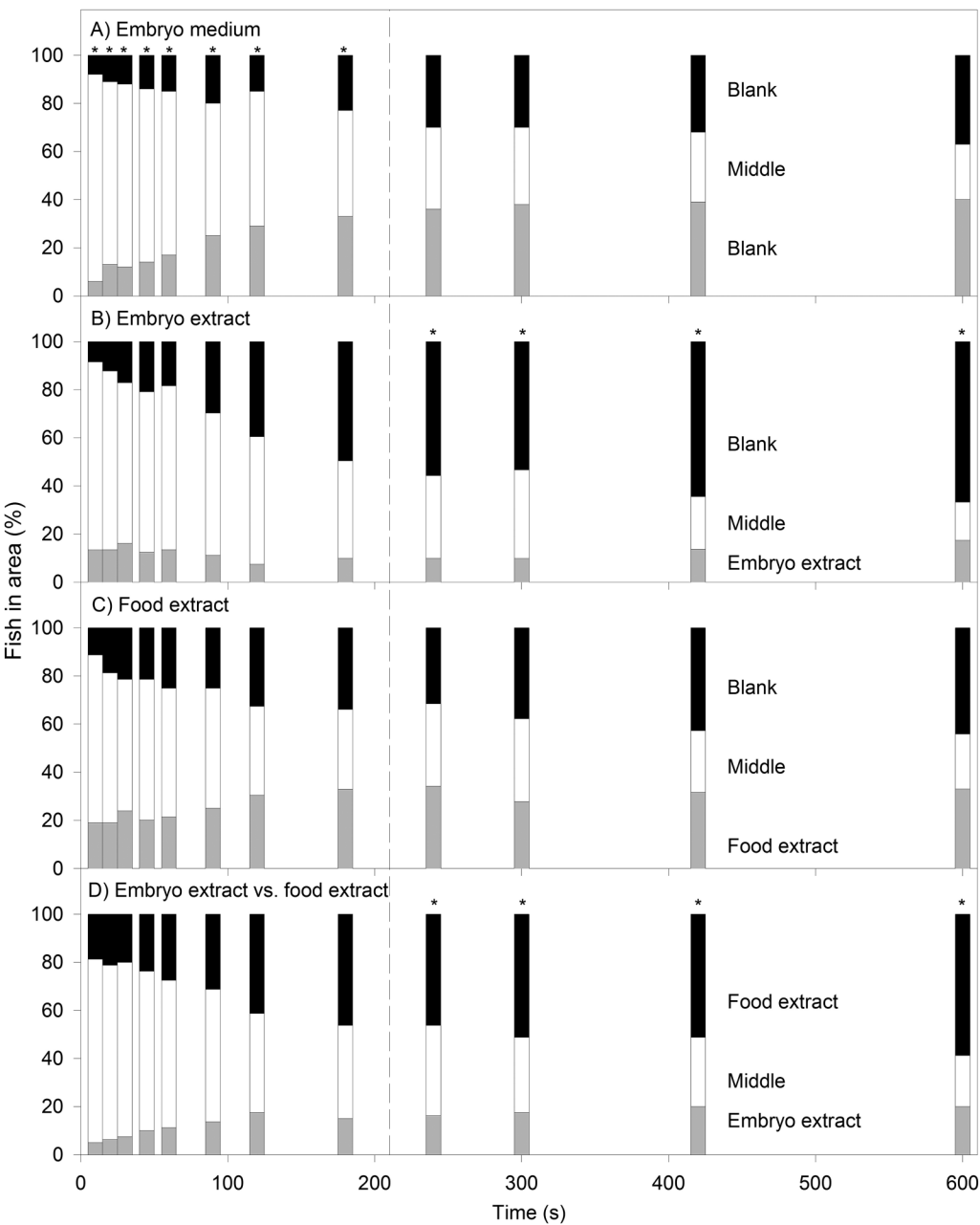


Fig. 1. Changes in occupancy of a three-chambered avoidance–attraction trough by zebrafish larvae (7 d post-fertilization). Embryos were acclimated to the central chamber and then barriers were removed to allow movement to the side chambers, which were of equal size. The number of larvae in each chamber was manually scored. Bars indicate the average number of embryos per chamber ($N = 8$ trials, with 9–11 embryos per trial). Shown are (A) responses to blank (embryo medium), (B) larval extract, (C) food extract, and (D) embryo extract vs. food extract. In (A), asterisks denote time points at which distributions were significantly different; in (B–D), asterisks denote a significant difference in the stimulus side vs. the distribution observed in control trials. The dashed line denotes the point of equal distribution of fish in control trials (see behaviour methods in the [Supplementary Material](#)).

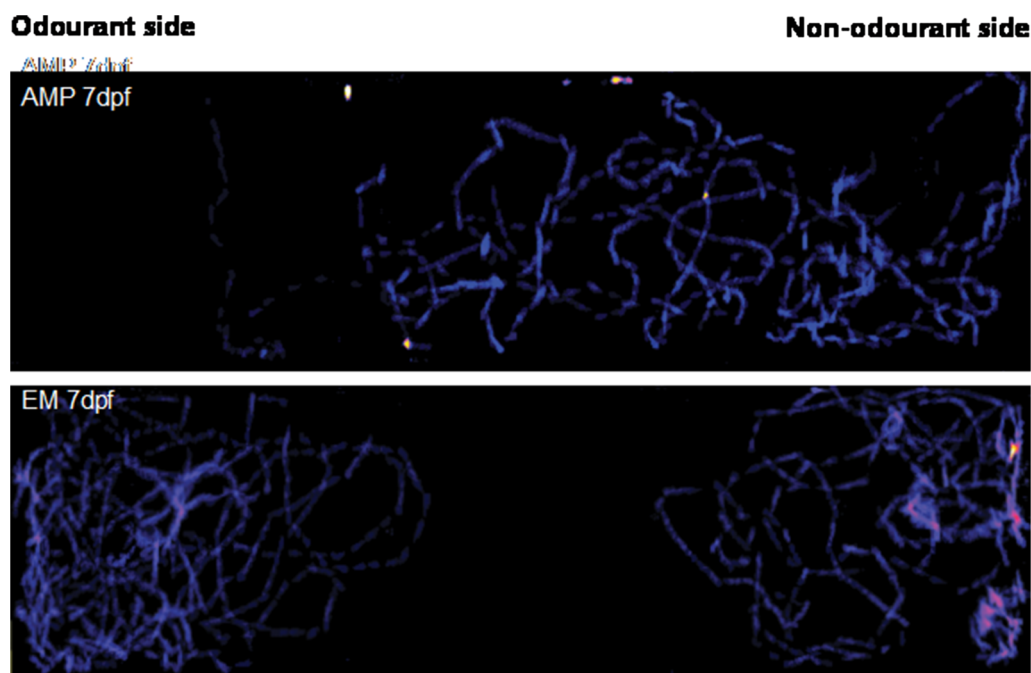


Fig. 2. Videogram analysis. Example videograms of movement of zebrafish larvae (7 d post-fertilization; $n = 9\text{--}11$) during exposure to adenosine monophosphate (AMP) or control (embryo medium; EM). Images represent 7–10 min during the trials. Image contrast has been modified to enhance appearance. The contrast was not altered during videogram analysis.

To determine stimulus attraction or avoidance, we examined the motion of all larvae considered together as the centre of mass over time. This was facilitated using videograms (**Fig. 2**). Videos were converted to AVI files (2 frames/s) using VirtualDub (General Public License) and then converted to videograms in ImageJ (National Institutes of Health, USA) using the protocol outlined by Wyeth et al. (2011). Average image stacks were created using every 10th frame between frame 100 and the last frame (rounded to the nearest hundred). The mean background image was imported into PhotoShop (Adobe, USA) and the Clone Stamp tool was used to eliminate stationary fish from the background image. Summed images were used to create videograms for the following time bins: 1–2, 2–3, 3–4, 4–5, 5–7, and 7–10 min (the first minute was not used owing to the water disturbance immediately following barrier removal). A maximum threshold of 20% was set for frame deletion within a time bin. As a result, some time bins have a smaller n -value than the total number of trials conducted per individual odourant. The shift in centre of mass was determined as the difference between the videogram centre of mass and the apparatus centre. Shifts were converted to a millimetre scale using the length of the apparatus. Data are presented as mean centre of mass shift \pm SEM.

Physiological responses

Olfaction

To determine whether nucleobases evoked responses at the olfactory epithelium, we recorded electro-olfactograms (EOGs), which capture neuron generator potential responses, from another model fish, the goldfish. We did not use zebrafish owing to challenges in measuring EOGs from such a small fish. We acknowledge that there may be species differences, but the goal was to provide

evidence that olfaction may play a role in nucleobase detection in fishes, and bony fishes are not dissimilar in olfactory abilities (Tierney 2015). Goldfish were given an induction dose of 200 mg/L tricaine methanesulfonate (Syndel, British Columbia) and a maintenance dose of 100 mg/L, perfused over the gills. The covering of the right naris was surgically removed and a recording electrode was placed on the third lamella from the right of the olfactory rosette; a reference electrode of the same composition was placed within the water bath. EOGs were taken as the maximum change from baseline and were recorded from the olfactory rosettes using Ag/AgCl electrodes embedded in 2% agar and 1 mol/L NaCl plugs in borosilicate capillary tubes, as described earlier (Evans and Hara 1985). Capillary tubes were pulled using an electrode puller to achieve a tip diameter of 25 μm , trimmed using forceps to achieve a diameter of 100 μm , and flame polished before being filled. All signals were AC coupled, filtered between 0.1 and 1000 Hz, and amplified 1000 \times . All recordings were acquired using a PowerLab 4/25 system and Chart5 software (ADInstruments, Colorado, USA). To evoke responses, we perfused the olfactory epithelium with background water for 10 min and delivered odourants in 2-s pulses at least 2 min apart. If goldfish did not have EOGs evoked by at least two odourants or if EOGs were <0.5 mV in magnitude, the electrode was repositioned. A maximum of three repositionings were allowed per fish; if conditions were not met on the third repositioning, the fish was not used.

Concentration–response curves were constructed for a subset of nucleobases including the purine nucleosides, guanosine and adenosine, and two purine derivatives, adenine and hypoxanthine, as previously described (Evans and Hara 1985). Each compound was tested at concentrations of 1×10^{-9} , 1×10^{-7} , and 1×10^{-5} mol/L, a range previously used to produce a concentration–response curve for amino acids (Friedrich and Korsching 1997). For each fish, three EOGs were recorded at each concentration, and the average of these was taken as the response. Responses to background water and to 1×10^{-5} mol/L L-serine were also evaluated. Responses to background water were recorded to detect whether any potential was being generated by the switching on of the solenoid valve controlling the perfusion mechanism. Responses to L-serine were recorded to demonstrate that fish were able to smell an odourant that has already been shown to evoke a response within the species. Cross-adaptation experiments were conducted as described earlier (Bruch and Rulli 1988). With cross adaptation, if a compound shares a common receptor with another, previous exposure to one compound will diminish the response to the other. Olfactory tissue was perfused with dechlorinated municipal tap water for 10 min before adenine, adenosine, hypoxanthine, or guanosine was perfused at a concentration of 1×10^{-5} mol/L. Adaptation was allowed for 30 min or until EOGs evoked by twice the concentration of the adapting odourant (2×10^{-5} mol/L) were the same as those of background water pulses before adaptation.

Gene expression

Zebrafish larvae were exposed to a mixture of ATP, AMP, adenosine, and adenine (1×10^{-5} mol/L) from 3 to 7 dpf. Control embryos were held in EM throughout the exposure period; all solutions were changed daily. These exposures are similar to the whole-organism exposures that could occur in behavioural trials, although they are longer. At 7 dpf, larvae were euthanized on ice, preserved in RNAlater[®] (Thermo Fisher, Waltham, Massachusetts, USA), and stored at -20°C until RNA extraction. Each sample of total RNA was extracted from 33 to 35 pooled whole larvae using TRIzol[®] Reagent (Ambion, Carlsbad, California, USA) according to the manufacturer's instructions. Extracted RNA was then purified using an RNeasy[®] Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol for RNA cleanup. Genomic DNA contamination was removed by a 30-min on-column incubation with DNase using an RNase-Free DNase Set (Qiagen).

Purified RNA was suspended in 20 μL of RNase-free water and stored at -80°C until analysis. RNA quality and concentrations were measured using a NanoVue spectrophotometer (GE Healthcare Life

Sciences, Chicago, Illinois, USA) and an RNA Nano 6000 Assay Kit for the Agilent 2100 Bioanalyzer (Agilent, Santa Clara, California, USA). RNA samples were sent to Genome Quebec for high-throughput RNA sequencing (RNA-seq) analysis. Sequencing was performed to a depth of 50 million reads per sample using an Illumina HiSeq 2000 system with a 2×100 -bp paired-end strategy and a 6-bp index read to facilitate library multiplexing (Baker and Hardiman 2014). Data were subjected to Illumina quality control procedures (>80% of the data yielded a Phred score of 30). Secondary analysis was performed using an OnRamp Bioinformatics Genomics Research Platform (OnRamp Bioinformatics, San Diego, California, USA) (Hardiman et al. 2016; Davis-Turak et al. 2017). OnRamp's advanced Genomics Analysis Engine utilized an automated RNA-seq workflow to process the data, including (1) data validation and quality control; (2) read alignment to the zebrafish genome (DanRer7) using TopHat2 (Trapnell et al. 2012), which revealed >78% mapping of the paired-end reads; (3) generation of gene-level count data with HTSeq; and (4) differential expression analysis with DESeq2 (Love et al. 2014), which enabled the inference of differential signals with robust statistical power (Genomics Research Platform with RNA-seq workflow v1.0.1, including FastQValidator v0.1.1a, Fastqc v0.11.3, Bowtie2 v2.1.0, TopHat2 v2.0.9, HTSeq v0.6.0, and DESeq v1.8.0).

The resulting Sequence Alignment Map files were sorted and input into the Python package HTSeq to generate count data for gene-level differential expression analyses. Transcript count data from DESeq2 analysis of the samples were sorted according to their adjusted p -value or q -value, which is the smallest false discovery rate (FDR) at which a transcript is considered significant. FDR adjustment is needed with large data sets such as those resulting from RNA-seq. FDR is the expected fraction of false positive tests among significant tests and was calculated using the Benjamini–Hochberg multiple testing adjustment procedure. Statistical analysis of pathways and gene ontology terms was performed using this sorted transcript list as described previously (Kozak et al. 2013; Paolini et al. 2014) and using the ToppGene Suite (Chen et al. 2009) and GOrilla (Eden et al. 2009). Sequencing data were submitted to Gene Expression Omnibus (National Center for Biotechnology Information) and given accession number GSE97016.

Statistics

Behaviour

Centre of mass data for each stimulus at each day were analyzed using a two-way (time \times stimulus) repeated-measures analysis of variance (ANOVA) followed by the Holm–Sidak post hoc test. Data sets that violated normality were rank transformed prior to parametric statistical analysis (Conover 1980). At 6 and 7 dpf, EM controls for odourant set 2 displayed a bias to the right side of the avoidance–attraction apparatus. As a result, the 6 and 7 dpf odourant trials from set 2 were compared with EM controls from set 1, where no bias was present. Values that statistically showed a bias to the right side were corrected prior to comparison with set 1 EM controls. The correction factor (CF) was defined as follows:

$$CF = \text{Mean EM shift odourant set 1} / \text{Mean EM shift odourant set 2}$$

Statistical analysis was performed and graphs were created using SigmaPlot 11.0 (Systat, California, USA). Significance was accepted at $p < 0.05$. All error bars are standard error of the mean.

Physiological responses

For EOG responses, concentration responses were analyzed using a repeated-measures (fish) one-way (concentration) ranked ANOVA followed by Tukey's post hoc test. Cross-adaptation analysis was performed in a similar manner, except that pre-exposure EOG values of each nucleobase were

compared with values during adaptation. For presentation purposes, changes in EOGs are given as percent change. Error was calculated as the average of the preadaptation and adapted responses, scaled by the magnitude of change.

RNA-seq analysis

Fold-change (FC) estimation and hypothesis testing for differential expression were performed using the DESeq2 Bioconductor library (Anders and Huber 2010; Anders et al. 2013; Love et al. 2014). For each gene, DESeq2 reported an estimated FC and provided an adjusted p - or q -value equivalent to the smallest FDR incurred when declaring that test significant.

Results

Behavioural responses

Overall, purines (double-ring nucleobases) induced behavioural responses, whereas pyrimidines (single-ring nucleobases) did not. When observed, behavioural responses were almost exclusively avoidance, and all tended to occur later in the trials (i.e., >5 min). Responses to un-phosphorylated compounds (adenine and adenosine) occurred early in development (5 dpf), while those to phosphorylated compounds (AMP and ATP) occurred later (7 dpf) (Fig. 3). Specifically, adenine was avoided at 7–10 min ($p = 0.048$), and a similar trend was present for adenosine at 7–10 min ($p = 0.056$). In contrast, larvae avoided AMP and ATP at 7 dpf, both at 5–7 min (AMP, $p = 0.069$; ATP, $p = 0.019$) and at 7–10 min (AMP, $p = 0.013$; ATP, $p = 0.004$). Hypoxanthine and guanine-based purines (those with a 6-oxo group on the major ring) did not evoke avoidance or attraction, with one exception: GMP evoked attraction at 3–4 min ($p = 0.029$) (Fig. 4). Interestingly, although H3NO did not evoke avoidance, in a flow-through chamber it evoked an increase in distance travelled during the first minute of exposure (H3NO vs. EM: $75.8\% \pm 31.8\%$ vs. $-16.1\% \pm 16.3\%$; $p = 0.0326$; Fig. S3), which coincided with an increase in maximum velocity (H3NO vs. EM: 262 ± 31.0 vs. 138 ± 31.0 mm/s; $p < 0.05$; Fig. S4). No other compounds evoked changes in activity.

The pyrimidines CMP and UMP did not evoke responses at any time during the 10-min exposure window in 5–7 dpf larvae, nor did they evoke responses when mixed with the purines AMP and GMP (Fig. 5).

There were some interactions between time and stimulus, in which the centre of mass tended to diverge over the 10-min trials (all statistical results are given in Table S3). This was noted for adenosine at 5 dpf ($F_{5,117} = 4.87$, $p < 0.001$), AMP at 6 dpf ($F_{5,104} = 4.61$, $p = 0.001$), ATP at 5 and 7 dpf ($F_{5,111} = 5.167$, $p < 0.001$; $F_{5,116} = 2.39$, $p = 0.044$), and GMP at 5 dpf ($F_{5,110} = 3.34$, $p = 0.009$). However, pairwise differences were found only for ATP at 7 dpf (Fig. 3L).

Considering behavioural responses over time alone, an interesting trend was apparent: fish generally moved from one side of the tank to the other over 10 min. This was apparent for hypoxanthine, H3NO, guanosine, and the nucleotide mixture at 6 dpf ($F_{5,112} = 4.87$, $p < 0.001$; $F_{5,106} = 9.42$, $p < 0.001$; $F_{5,105} = 4.74$, $p < 0.001$; $F_{5,106} = 2.69$, $p = 0.027$) and for CMP and UMP at 7 dpf ($F_{5,116} = 6.83$, $p < 0.001$; $F_{5,117} = 7.48$, $p < 0.001$).

Physiological responses

Olfaction

Of the four purines tested, three evoked olfactory tissue responses with some degree of concentration dependency (adenosine, hypoxanthine, and guanosine; p -values: < 0.001 , 0.047, and < 0.001), while one did not (adenine, $p = 0.531$) (Fig. 6).

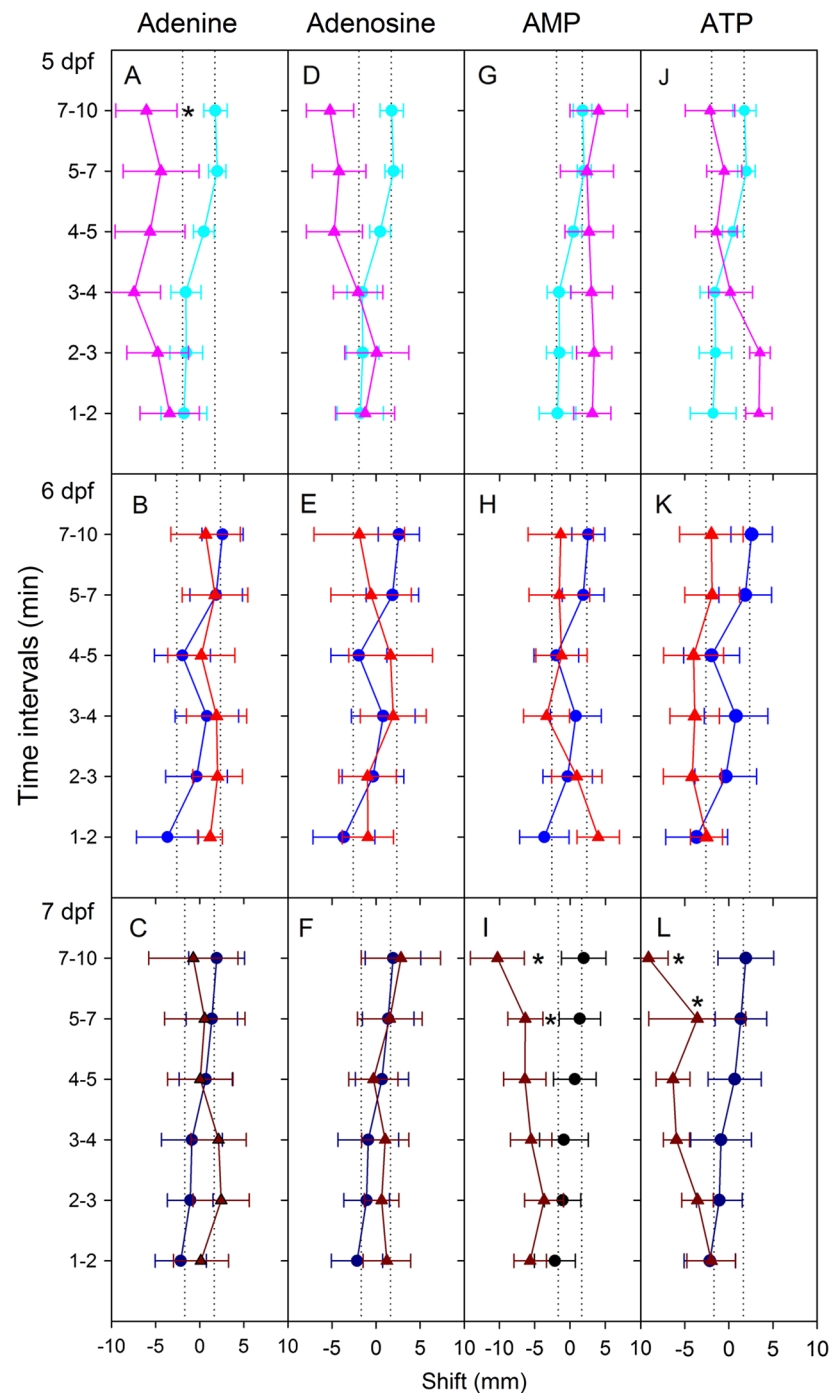


Fig. 3. Avoidance-attraction responses of zebrafish larvae to adenine- and adenosine-based purines. Movement was measured as a shift in the centre of mass. Shown are post-stimulus time interval bins at 5–7 d post-fertilization (dpf). All stimuli (triangles) were given at a final dilution of 1×10^{-5} mol/L. $N = 8-10$ trials, each with 10 larvae. A shift to the left indicates avoidance and a shift to the right indicates attraction; dotted lines represent the 95% confidence intervals for embryo medium (EM; circles). An asterisk denotes a significant difference from the EM shift. AMP, adenosine monophosphate; ATP, adenosine triphosphate.

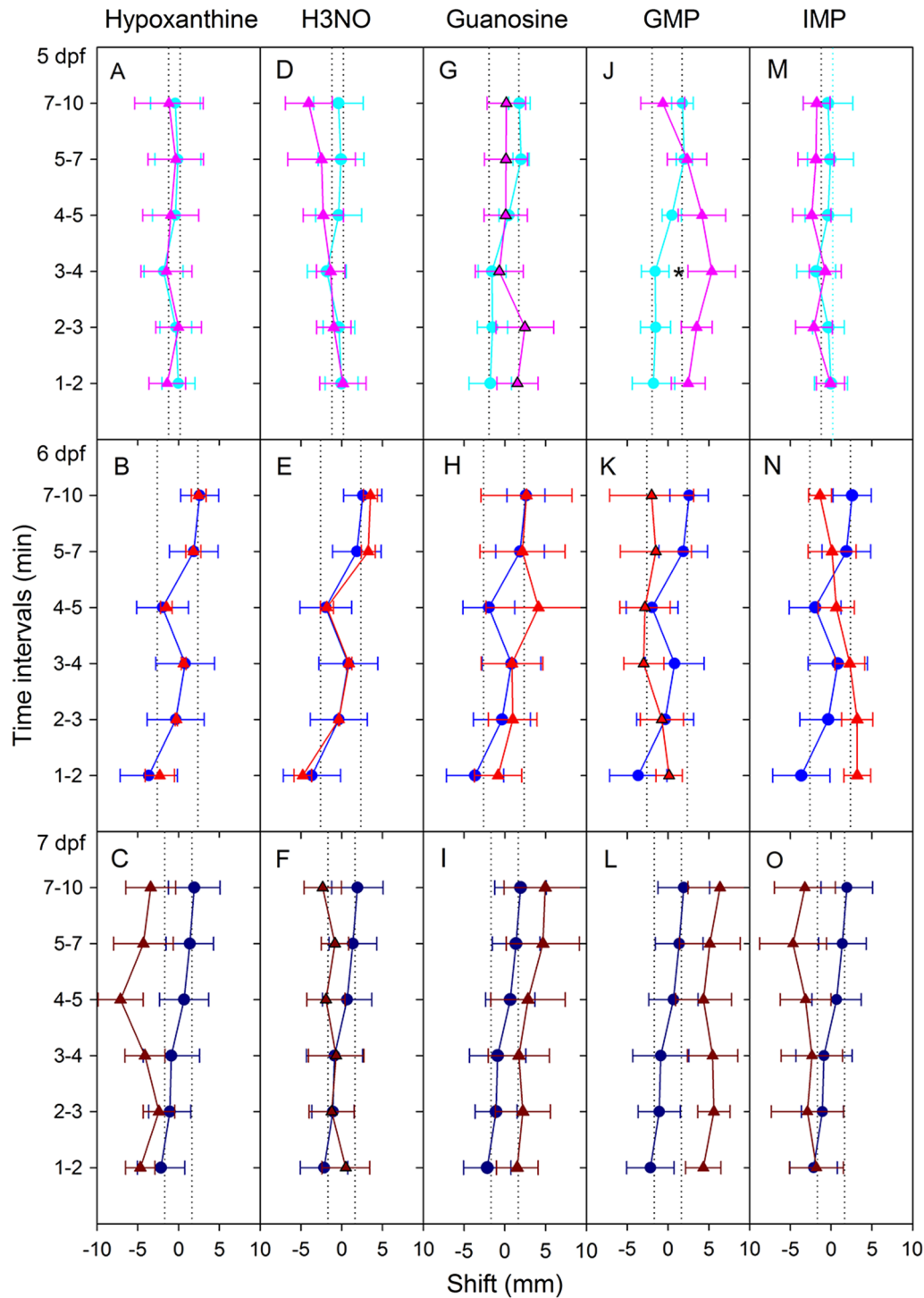


Fig. 4. Avoidance-attraction responses of zebrafish larvae to hypoxanthine- or guanine-based purines. Movement was measured as a shift in the centre of mass. Shown are post-stimulus time interval bins at 5–7 d post-fertilization (dpf). All stimuli (triangles) were given at a final dilution of 1×10^{-5} mol/L. $N = 8-10$ trials, each with 10 larvae. A shift to the left indicates avoidance and a shift to the right, attraction; dotted lines represent the 95% confidence intervals for embryo medium (EM; circles). An asterisk denotes a significant difference from the EM shift. H3NO, hypoxanthine 3-*N*-oxide (alarm compound); GMP, guanosine monophosphate; IMP, inosine monophosphate.

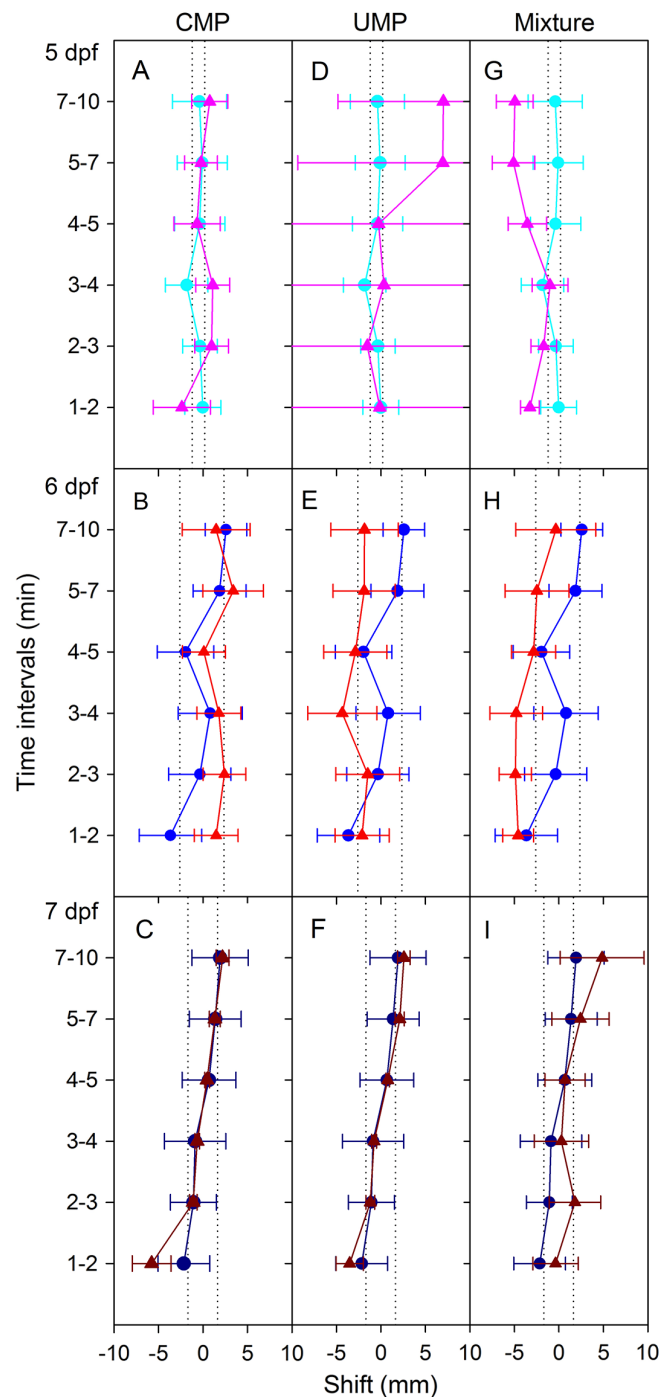


Fig. 5. Avoidance-attraction responses of zebrafish larvae to two pyrimidines and a mixture of purines and pyrimidines (adenosine monophosphate (AMP), guanosine monophosphate (GMP), cytosine monophosphate (CMP), and uracil monophosphate (UMP)). Embryo movement was measured as a shift in the centre of mass. Shown are post-stimulus time interval bins at 5–7 d post-fertilization (dpf). All stimuli (triangles) were given at a final dilution of 1×10^{-5} mol/L, except for the mixture, which was 4×10^{-5} mol/L. $N = 8-10$ trials, each with 10 embryos. A shift to the left indicates avoidance and a shift to the right, attraction; dotted lines represent the 95% confidence intervals for embryo medium (EM; circles). An asterisk denotes a significant difference from the EM shift.

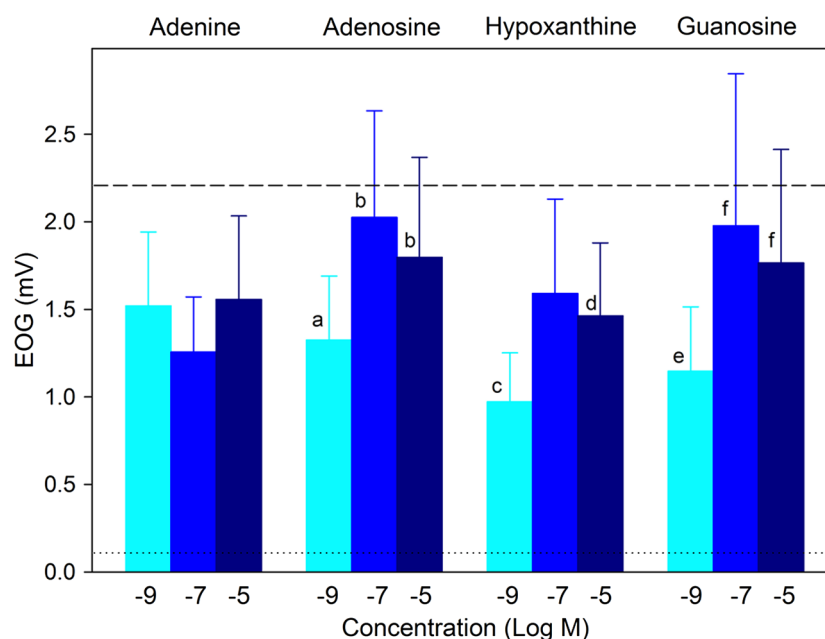


Fig. 6. Responses of olfactory epithelium evoked by increasing concentrations of purine nucleosides and recorded as electro-olfactograms (EOGs). Differences in EOGs within a purine are denoted by different letters. The dotted line indicates responses to background water; the dashed line indicates responses to 1×10^{-5} mol/L L-serine.

With cross adaptation, reciprocal knockdown of generator potential responses suggests a common receptor; a lack of reciprocal knockdown has less certain meaning. The former occurred for adenine and guanosine (adenine adaptation reduced guanosine responses, $p = 0.021$; guanosine adaptation reduced adenine responses, $p = 0.018$; Fig. 7). This was quite unexpected owing to appreciable structural differences between the two (see Table 1). The latter occurred with adenosine and hypoxanthine: adaptation to adenosine knocked down hypoxanthine responses ($p = 0.013$), but hypoxanthine adaptation did not affect adenosine responses. Conceivably, adenosine may have activated many hypoxanthine receptors, but hypoxanthine may not have activated many adenosine receptors. Alternatively, if we consider that the pathways may not be wholly separate, then perhaps adenosine activated the hypoxanthine pathway, but hypoxanthine did not activate the adenosine pathway.

Gene expression

Sequencing analysis identified 680 significantly differentially expressed genes (DEGs) at FDR <0.1 and 2045 DEGs at FDR <0.4, when referenced to the *D. rerio* genome (43 DEGs with a linear FC >2). These were analyzed using *GORilla*. The protein FASTA sequences from Ensembl for zebrafish were compared using Ensembl's homology to create protein FASTA files that contained a human Entrez gene ID that mapped to zebrafish (Renaud et al. 2017). This enabled a more comprehensive pathway-based analysis using human gene IDs with the ToppGene Suite. This analysis returned 1955 DEGs at FDR <0.1 and 4218 DEGs at FDR <0.4 (77 DEGs with a linear FC >2). The majority of the top 10 DEGs in both analyses belong to the tubulin, ubiquitin, and ribosomal protein gene families (Table S1).

Many zebrafish genes involved in nucleotide, guanosine triphosphate (GTP), and ATP binding were upregulated by exposure to the mixture of ATP, AMP, adenosine, and adenine (Table 2). The human

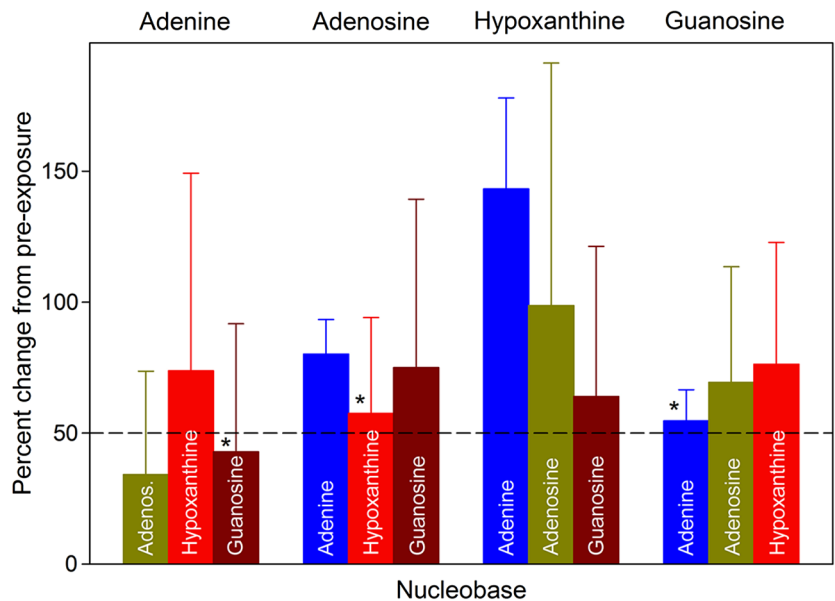


Fig. 7. Olfactory responses following previous stimulus exposure (cross adaptation). Shown are changes in pre- vs. post-adaptation electro-olfactogram (EOG) responses; reduced responses suggest that the nucleobases share one or more receptors (adenine, $n = 8$; adenosine, $n = 6$; guanosine, $n = 6$; hypoxanthine, $n = 6$). An asterisk denotes a difference from the preadaptation value; a dashed line indicates a 50% EOG reduction.

gene ID appended output indicated that genes involved in nucleotide binding, specifically guanine nucleotides, were upregulated by exposure (Table 3).

The top five altered molecular functions were very similar between the zebrafish and the human gene ID appended results, with the top four being the same (Table S2). These altered functions are mostly involved in cell component structure and RNA binding. Although the human gene ID appended analysis revealed many functions involving purine binding and transporter activity within the top

Table 2. Selected upregulated zebrafish genes involved in nucleotide, guanosine triphosphate (GTP), and adenosine triphosphate (ATP) binding.

Symbol	Description/function	Linear fold change	Adjusted <i>p</i> -value
zgc:153713	Ras homolog gene family, member A/GTPase protein	6.33	2.68E-65
zgc:55461	Tubulin, beta 2b/GTP binding	3.41	6.58E-47
zgc:86709	ATP binding	2.56	2.51E-15

Table 3. Selected genes involved in purine binding that were upregulated in the humanized output.

Symbol	Description	Linear fold change	Adjusted <i>p</i> -value
GNG13	Guanine nucleotide binding protein (G protein), gamma 13	2.36	1.09E-33
GBP6	Guanylate binding protein family, member 6	1.81	6.78E-24
GNB2L1	Guanine nucleotide binding protein (G protein), beta polypeptide 2-like 1	1.52	2.93E-11

Table 4. Selected altered gene ontology molecular functions from the humanized analysis (all within the top 50 altered functions) using ToppGene.

Name	Adjusted <i>p</i> -value
Guanyl ribonucleotide binding	8.56E-13
Guanyl nucleotide binding	9.06E-13
Purine ribonucleoside triphosphate binding	3.25E-9
Purine ribonucleoside binding	4.13E-9
Purine nucleoside binding	4.36E-9
Purine ribonucleotide binding	7.88E-9
Purine nucleotide binding	1.03E-8
Adenine transmembrane transporter activity	3.90E-4
Purine nucleobase transmembrane transporter activity	1.91E-3

50 altered functions (Table 4), when referenced to the zebrafish genome, adenosine receptor binding was the only function involving purine binding and transporter activity within the top 50 DEGs ($p = 0.01$).

Discussion

Purine nucleobases evoked behaviours, caused neuronal responses, and evoked changes in gene expression. Although some of the responses were expected, such as ATP avoidance, many were not, including neuronal responses at a very low threshold and the expression of many genes with unclear relevance to nucleobase exposure or chemosensation.

Behaviour

Apart from four instances among 36 different time–odourant combinations, larval zebrafish did not respond to nucleobases. When responses were observed, nucleobases evoked avoidance (with only one exception), and the responses were dependent on nucleobase structure and embryo age. The data demonstrate that adenine-containing compounds may cause behavioural change depending on dpf. Recently, it was demonstrated that ATP may be enzymatically converted to adenosine at the olfactory epithelium and that adenosine activates a novel receptor (Wakisaka et al. 2017). Given that we noted changes in response to adenine, ATP, and AMP, it is possible that adenine is capable of stimulating the same receptor or that it has its own unique receptor. The first option may be unlikely, as Wakisaka et al. (2017) found that adenine did not lead to activation of the same glomerulus in the olfactory bulb that adenosine did. Regardless, our findings are in agreement with previous studies that showed that adenine-containing compounds evoked the most potent responses (Friend and Smith 1971; Carr et al. 1986).

Attractive responses to ATP and AMP were expected owing to the ability of these compounds to evoke feeding in other species (Friend and Smith 1971; Smith and Griend 1976; Mearns et al. 1987; Miyasaki and Harada 2003; Romero and Schal 2014). However, such responses were not observed in zebrafish larvae, likely because of fish age. A study of feeding on nucleotide-enriched agar showed that nucleotides deterred feeding in Atlantic salmon parr but stimulated feeding in adult rainbow trout (Mearns et al. 1987). This raises the question, is there a reason why younger fish are deterred by nucleotides? There are at least two potential explanations: (1) stimulus naivety and (2) stimulus

significance. In the case of salmon parr, the fish have not yet made their oceanic migration and it is possible that the nucleotides used in the study (sourced from shrimp) were not yet familiar as food odourants. The same is possible for zebrafish larvae—they have not yet been exposed to food and therefore may not associate nucleotides with food. In support of this, adult zebrafish were attracted to adenosine-containing compounds (Wakisaka et al. 2017). Alternatively, considering that nucleotides are found in biological tissues such as blood, perhaps salmon parr and zebrafish larvae are prey (Mearns et al. 1987; Carr et al. 1996; Engeszer et al. 2007; Romero and Schal 2014), and so ATP indicates an alarm cue (e.g., recent release from a nearby sibling).

A relationship between the number of phosphates and the strength of the behavioural response was not observed. In studies of insects, decreased phosphorylation was associated with decreased stimulatory efficacy (Friend and Smith 1971; Smith and Griend 1976; Romero and Schal 2014). In the present study, there was no observed relationship between the number of phosphates and the capacity to evoke behaviours. However, we did not directly compare responses to AMP and ATP and so cannot draw definitive conclusions. To confirm our finding, future studies should include dual odourant trials (i.e., comparison of two odourants, one on each side of the attraction–avoidance apparatus).

We were surprised that no pyrimidines evoked responses, as cytidine- and uridine-containing nucleotides initiated gorging behaviour in the kissing bug, and CTP stimulated spiny lobster chemosensory cells (Friend and Smith 1971; Smith and Griend 1976; Carr et al. 1996). Based on these studies, we expected that the phosphate groups in nucleotides would drive the behavioural responses. We found the opposite, as it appears that for embryonic zebrafish, nucleobase structure is the determining factor. Interestingly, in abalone and oriental weatherfish (*Misgurnus anguillicaudatus* (Cantor, 1842)), neither CMP nor UMP increased feeding responses; however, the pyrimidine thymidine was effective (Brown et al. 2001). We suggest that pyrimidine responses be tested in adult zebrafish.

It was also surprising that H₃NO, a potent alarm-inducing compound, did not evoke avoidance. However, when zebrafish larvae were placed in a flow-through chamber and forced to experience H₃NO, they responded rapidly (≤ 1 min) with increased swimming activity and speed. Typically in adult fishes, alarm response is apparent in dashing, which may be followed by immobility or “freezing”. However, in adult zebrafish, H₃NO has only been observed to evoke increases in activity (Speedie and Gerlai 2008; Parra et al. 2009; Mathuru et al. 2012; Kalueff et al. 2013; Gallus et al. 2016). Our study agrees with the adult data and demonstrates that alarm-like behaviour can be evoked by H₃NO just days into zebrafish development.

Hypoxanthine did not evoke avoidance, which confirms earlier work in fathead minnows (*Pimephales promelas* Rafinesque, 1820) and finescale dace (*Chrosomus neogaeus* (Cope, 1867)), in which compounds that lacked a nitrogen-oxide functional group, including hypoxanthine, failed to evoke alarm responses (Brown et al. 2000). The results of the present study suggest that the nitrogen-oxide group is likely related to an alarm response and the basic purine ring structure is related to an avoidance response.

The nucleotide mixture did not evoke any avoidance or attraction response despite containing behaviourally active AMP and GMP. This may be a result of the ratio and combination of compounds within the mixture, or it may be because the active stimuli were lost in the noise of the inactive. Because nucleotides are most likely to appear in nature as mixtures, additional combinations should be tested that better represent the nucleotide composition of food and conspecific tissue extracts.

There was just one example of zebrafish attraction response—to GMP—and it was a true oddity because it occurred 3–4 min into the trial (no other responses were seen at this time). In contrast to GMP, guanosine evoked no response, suggesting that the presence of phosphate has the potential to alter odourant efficacy. However, because not all of the tested nucleotides evoked a response

(UMP and CMP did not), the presence of phosphate alone does not appear sufficient to evoke a behavioural response. This leaves three possibilities: (1) odourant–receptor interaction is based on three-dimensional structure and is therefore influenced by the complete form of the nucleotide, (2) GMP circumvents the odourant–receptor interaction, or (3) this finding was a statistical aberration.

Olfactory-evoked behaviours vary with age

Odourant responses were observed almost exclusively at 5 and 7 dpf, not 6 dpf. This is interesting because previous studies have shown that zebrafish develop kin odour recognition at 6 dpf (Gerlach et al. 2008; Hinz et al. 2013). At 6 dpf, zebrafish larvae exhibit an increase in thyroid hormone receptor β mRNA and whole-body thyroxine (Liu and Chan 2002; Chang et al. 2012). In a salmonid species (*Oncorhynchus kisutch* Walbaum, 1792) known to imprint to the odours of the natal stream, elevated plasma thyroxine was associated with olfactory epithelium proliferation (Lema and Nevitt 2004). This behavioural and hormonal evidence has led to the theory that olfactory imprinting occurs at 6 dpf. If imprinting occurs at 6 dpf, it is possible that the dynamic status of the olfactory system at this time affects odourant interpretation. An alternative explanation for temporal changes in responses may be asynchronous expression of olfactory receptors, as studies have shown that the onset of expression of odourant receptors varies (Barth et al. 1996; Argo et al. 2003).

Physiological responses to nucleobases

Olfactory responses

In the current study, response at the olfactory epithelium should only be used to address three questions: (1) is the response to nucleobases similar to the response to known odourants (is there a concentration–response relationship), (2) is nucleobase sensitivity similar to sensitivities to known odourants, and (3) do nucleobases share receptors (does the presence of one diminish the response to another)? For a variety of reasons, drawing direct comparisons between behavioural responses of zebrafish larvae and olfactory responses of goldfish should be done with caution. Among the reasons are differences between species, developmental stages, and importantly, nucleobase exposure routes. Zebrafish larvae may be receiving nucleobase sensory input from numerous routes (e.g., the gustatory system (Hansen et al. 2002; Yacoub and Browman 2007; Oka and Korsching 2011) or solitary chemosensory cells (Kotrschal et al. 1997)), whereas goldfish responses were recorded from isolated olfactory epithelium. At a minimum, goldfish olfactory epithelial responses suggest that nucleobases may serve as odourants and, since adenine, adenosine, and guanosine tended to knock down responses to each other, that some nucleobases share common receptors. Recently, it was found that ATP is converted to adenosine in a three-step, two-enzyme system on the olfactory epithelium and that adenosine acts on a specific receptor (Wakisaka et al. 2017).

Some of the nucleobase-evoked responses (i.e., those to adenosine, hypoxanthine, and guanosine) were consistent with concentration dependency, as neuron generator potential responses increased between 1×10^{-9} and 1×10^{-7} mol/L. This suggests that zebrafish behavioural responses may have been partly due to olfaction. What was curious, however, was that the olfactory responses occurred between 1×10^{-9} and 1×10^{-7} mol/L and not between 1×10^{-7} and 1×10^{-5} mol/L. We selected the concentration range to capture responses similar to the sigmoidal responses to amino acids, which begin at approximately 1×10^{-8} mol/L and plateau at 1×10^{-3} mol/L (Hara and Zhang 1996; Friedrich and Korsching 1997; Baldwin et al. 2003; Tierney et al. 2007). Because robust responses were present at nucleobase concentrations of 1×10^{-9} mol/L, the olfactory epithelium appears to be more sensitive to nucleobases than amino acids. In support of this, Parra et al. (2009) found that H3NO evoked alarm responses at 1×10^{-9} mol/L. Olfactory responses at such high dilution are not unusual; rather, they are common to odourants considered “pheromones” (Rosenblum et al. 1991; Moore and

Waring 1996; Stacey 2003). Responses to pheromones may occur at dilutions as high as 1×10^{-12} mol/L (Li et al. 1995; Keller-Costa et al. 2014). We are not suggesting that nucleobases are pheromones (i.e., species-specific signals), but we do leave open the possibility that nucleobases are effective at high dilution and may evoke physiological responses.

Intriguingly, responses to adenine were independent of concentration over a large (10 000-fold) range. Perhaps adenine does not stimulate responses through an olfactory pathway. Olfactory neurons have cyclic nucleotide-gated (CNG) channels that open in response to elevated cyclic nucleotides (cAMP and cGMP), allowing for the influx of calcium and subsequent depolarization of the neuron (Goulding et al. 1992; Zheng and Zagotta 2004). Embryonic zebrafish begin expressing CNG cation channels in the olfactory placode at approximately 24 h post-fertilization.

Gene expression

Gene expression data revealed numerous DEGs following exposure to the mixture of ATP, AMP, adenosine, and adenine. Many genes and biological processes involved in purine binding, specifically guanine binding, were upregulated by the exposure, which was expected because the mixture consisted of purines. The upregulation of genes involved in GTP binding suggests increased cell signaling, which was not unexpected (Mombaerts 1999; Howard et al. 2001). G proteins are also involved in regulating the actin cytoskeleton, which may explain the upregulation of cytoskeleton-related genes (Matsumoto et al. 2004).

Although numerous genes encoding G proteins were upregulated by the exposure, none have been specifically associated with the recognition of exogenous purines. Many of the GTP-binding genes that were altered have not been fully characterized. These genes (and proteins) should be further studied to elucidate any involvement in exogenous nucleotide binding and chemosensation.

The human gene ID appended analysis of the zebrafish DEGs supported the results obtained from the zebrafish genome. Both sets of results revealed many of the same altered genes and molecular functions. Remarkably, the humanized mRNA analysis predicted that the larvae had been exposed to guanosine, which is similar to the exposure mixture used in this study.

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Author contributions

ALS, BJB, and KBT conceived and designed the study. ALS, BJB, DDL, MQW, AG, CRL, and KT performed the experiments/collected the data. ALS, BJB, DDL, MQW, AG, CRL, KT, ESH, GH, and KBT analyzed and interpreted the data. KT, ESH, GH, and KBT contributed resources. ALS, BJB, DDL, CRL, KT, ESH, GH, and KBT drafted or revised the manuscript.

Competing interests

The authors have declared that no competing interests exist.

Data accessibility statement

All relevant data are within the paper, the Supplementary Material, and in the Gene Expression Omnibus (NCBI) (ncbi.nlm.nih.gov/geo/; accession number for sequencing data is GSE97016).

Supplementary material

The following Supplementary Material is available with the article through the journal website at doi:[10.1139/facets-2017-0101](https://doi.org/10.1139/facets-2017-0101).

Supplementary Material 1

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