

Supplemental materials for:

Using transcriptomics to examine the physiological status of wild-caught walleye (*Sander vitreus*)

Jennifer D. Jeffrey, Matt J. Thorstensen, Eva Enders, Jason R. Treberg, and Ken M. Jeffries

Methods

Differential expression analysis

Salmon v0.13.3 (Patro et al. 2017) was used to estimate transcript abundances using a previously assembled reference transcriptome for walleye generated by Jeffrey et al. (2020) (Sequence Read Archive Accession SRP150633). The R/Bioconductor package “tximport” (Soneson et al. 2016) was used to estimate counts from abundances at the gene-level that were scaled using the average transcript length, averaged over samples, and library size (i.e., argument `countsFromAbundance = lengthScaledTPM`). Differential expression of genes was examined using the R/Bioconductor package “edgeR” (Robinson et al. 2010). Only genes with at least one count per million across eight samples were considered for further analysis, representing 34,185 genes, 12.9% of genes, and $> 98.0 \pm 1.9\%$ (mean \pm standard deviation) of reads per sample. An effective library size was calculated using “`calcNormFactors`” in “edgeR” to normalize library sizes across samples (Robinson and Oshlack 2010). A general linear model was run in “edgeR” with sampling site and year as factors, and quasi-likelihood *F*-tests were used for hypothesis testing (Lun et al. 2016). *A priori* contrasts were designed to compare sampling sites (i.e., Dauphin River vs. Red River, Dauphin River vs. Matheson Island, Matheson Island vs. Red River) within a year and genes were considered differentially expressed at a Benjamini-Hochberg corrected False Discovery Rate (FDR) < 0.05 .

To identify differentially expressed genes that were specific to each sampling site, lists were generated for each sampling site and consisted of genes that were differentially expressed at that site compared to both other sites within a given year (e.g., Red River vs. both Matheson Island and Dauphin

River; Matheson Island vs. both Red River and Dauphin River; Dauphin River vs. both Red River and Matheson Island). The site-specific differentially expressed genes were then identified as being consistently different across years, different only within 2017, or different only within 2018. Annotation information for differentially expressed genes was retrieved from the previously annotated walleye reference transcriptome (Jeffrey et al. 2020). Differentially expressed genes that did not have available annotation information from the reference transcriptome or were identified as uncharacterized or non-vertebrate, were re-blasted using blastx and the non-redundant protein sequences database. Any non-vertebrate genes were removed prior to gene enrichment analysis.

Statistical analysis for selecting candidate genes for qPCR

Using the RNA-seq data, a set of 20 candidate genes were selected for qPCR using a strategy similar to that of Akbarzadeh et al. (2020) to develop qPCR assays for hypoxia biomarkers in salmonids.

Annotated genes representing consistent differences among sampling sites, across years were subjected to a PCA using the scaled and normalized cpm values and the “FactoMinerR” package in R. The PC associated with the separation of sampling locations was identified and the genes most correlated to this PC axis were determined using the “dimdesc” function in “FactoMinerR”. Candidate genes were selected based on their strong correlation with the PC that was associated with the separation of locations ($p < 0.05$) that were also not significantly correlated with the PC that separated years ($p > 0.1$). To verify separation of locations for the selected genes and to make comparisons to the qPCR data, an additional PCA using only the logCPM values for the 20 candidate genes was run. The PC representing separation by location was identified, and a two-way ANOVA was used to examine the fixed factors of location, year, and location \times year for the PC scores of this axis. Residuals were examined for normality and equal variance and significant effects were further explored using a Tukey’s HSD post-hoc test as above.

Comparison of RNA-seq data with qPCR

In addition to the Spearman's correlation analysis used to compare the RNAseq and qPCR log₂ fold changes for the 20 candidate genes, additional analyses were carried out to establish whether the individuals examined in the RNA-seq study were representative of the larger sample size assessed using qPCR. Using only individuals from Red River, Matheson Island, and Dauphin River (i.e., analyzed across both platforms), a linear model was developed for each target gene with location, year, the interaction of location and year, the platform, as well as total length and mass as fixed factor. A stepwise Akaike Information Criterion (AIC) was used to compare models and determine the best fit for the data. The "best fit" model was run, and the residuals were examined for normality and equal variance using the "check model" function from the R package, "performance". If the data did not meet the assumptions of the linear model, the data were rank transformed (only for *actn4*). Significant effects of categorical factors were further explored with a Tukey's HSD post-hoc test using the "emmeans" package in R. There were no significant effects of total length or mass when these factors were included in the models.

Results

Enrichment analysis for Matheson Island differentially expressed genes

Fewer genes (180 total and 168 unique) were differentially regulated in walleye from Matheson Island compared to Red River and Dauphin River (Tables S3, S4), and significant enrichment of GO terms were only evident in 2018. In 2018, GO terms related to the immune response were enriched for genes up-regulated in Matheson Island fish compared to fish from the other two locations (Fig. S3a), and included genes such as *tyrosine-protein kinase Lck (lck)* and *ZAP-70 (zap70)* and *lymphocyte cytosolic protein 2 (lcp2)* (Table S5). For genes down-regulated in fish from Matheson Island in 2018, the 'endoplasmic reticulum lumen' GO term was significantly enriched (Fig. S3b), involving genes such as *cytoskeleton-associated protein 4 (ckap4)* and *transport and Golgi organization protein 1 homolog*

(*mia3*) (Table S5). Significant enrichment of GO terms for genes of Matheson Island walleye that were down-regulated compared to Red River fish and up-regulated compared to Dauphin River fish in 2018 included GO terms related to the respiratory electron transport chain (Fig. S3c) and mitochondrial genes associated with complex I (*mt-nd3-6*, *mt-nd4l*) and complex III (*mt-cyp*) (Fig. 5; Table S6).

Table S1. Metadata for walleye (*Sander vitreus*) collected from Red River, Matheson Island, and Dauphin River in the Lake Winnipeg system included in the RNA-sequencing analysis.

Location	Year	Total <i>n</i>	Female <i>n</i>	Unknown sex <i>n</i>	Total length (mm)	Mass (kg)
Red River	2017	8	8	0	666 ± 42	3.29 ± 0.67
	2018	8	8	0	662 ± 47	3.68 ± 1.01
Matheson	2017	8	8	0	613 ± 57	2.30 ± 0.65
	2018	8	7	1	650 ± 65	2.80 ± 1.07
Dauphin River	2017	8	8	0	578 ± 61	1.97 ± 0.40
	2018	8	5	3	555 ± 57	1.83 ± 0.68

Data are presented as means ± sd.

Table S2. Oligonucleotide primers for qPCR in walleye (*Sander vitreus*).

Gene name	Protein name	Forward primer (5'–3')	Reverse primer (5'–3')	Product (bp)	Eff. (%)
<u>Cytoskeleton organization</u>					
<i>actn4</i>	Alpha-actinin-4	CTCTGCGAAAGAGGGTCTTCTC	GACCGTCCTTCCAGCTAATGTG	98	102
<i>actr2</i>	Actin-related protein 2	ACCCGATGGAGAACGGTATG	GGTCGGGACCAAAGGTGTAGT	75	107
<i>arf6</i>	ADP-ribosylation factor 6	ATCCTGATCTTCGCCAACAAA	GGCCTAGCTTCTCCTGGATCTC	73	99
<i>arhgdia</i>	Rho GDP-dissociation inhibitor 1	GCGGCACCTACACCATCAA	CACCAATCCCAGGAGAGATGA	70	104
<i>cdc42</i>	Cell division control protein 42 homolog	AGACAGCAACACGATCGAGAAG	TGAGCTCACGAGCCAGCTT	86	99
<i>cot11</i>	Coactosin-like protein	CCTGGATCGGTGAGAACATCA	CTTTGACCAGCGCCTTGTC	69	103
<i>fam49b (cyrib)</i>	Protein FAM49B (CYFIP-related Rac1 interactor B)	GCACCTGGAGCAGAAACAGTCT	GCAGCGTGAAGTGCAGGAT	62	104
<i>pfn2</i>	Profilin-2	CAGCTACTGCATGCACCTGAA	GCCTTGCCGACACAGATGT	66	99
<u>Stress response</u>					
<i>akr1a1b</i>	Aldo-keto reductase family 1 member A1-B	AGCAGTTATTTGGGCATTGGA	CCAAATGTCTCATGCAGTGCTT	102	91
<i>dnajc9</i>	DnaJ homolog subfamily C member 9	TACGAGGTGCTCGGCATCA	TGGACTTTCAGCGACACTTTGT	80	94
<i>gpx1</i>	Glutathione peroxidase 1	CATGAGCGGTACACCAGCAA	TTCTCCTGGTGGCCGA ACT	71	91
<i>slc25a24</i>	Calcium-binding mitochondrial carrier protein SCaMC-1	GGAGGGAGCATTTTCTGTTTAA	CACCAATATCCAGCACCGTAGA	87	101
<i>ube2j1</i>	Ubiquitin-conjugating enzyme E2 J1	TCATCCAGACAGGCAAGTGAGA	TGCAGACGTGGAGGTGTCTTC	81	107
<u>DNA repair</u>					
<i>rpa2</i>	Replication protein A 32 kDa subunit	TCCGGGCACGTATGTCAAA	GCCACGATAGATCGGTGGTT	66	97

<u>Golgi protein transport</u>					
<i>ap1s1</i>	AP-1 complex subunit sigma-1A	CAGGGAGCTGATGCAGATAGTG	GGTCCCTCCATTCGAGGAA	68	100
<i>eipr1</i>	EARP-interacting protein homolog	AAGTTCACCTCGGGCAAGTG	CTCGTATGGCCGTGTCATTG	76	104
<u>Immunity</u>					
<i>tnfaip8l2</i>	Tumor necrosis factor, alpha-induced protein 8- like protein 2	CACCAAAAGCTCTGTCCAAATG	GCGGTACAGTTCGTCCAGGAT	67	104
<u>Ion transport</u>					
<i>mcu</i>	Calcium uniporter protein, mitochondrial	TGCTTACACGCCAGGAGTATGT	CTTTTCACCCCCTTGTGGAA	79	95
<i>cnm4</i>	Metal transporter CNNM4	GGTGTCATGGCACTCAATGC	GGTTCAACCCGCTGACGTTA	67	106
<u>Signaling pathways</u>					
<i>dgka</i>	Diacylglycerol kinase alpha	GCTGCACCTGTTGCAAGTACA	GCGACAACCAATCTTCGGTTT	104	107

Table S3. Number of differentially regulated genes with annotation information, the number of unique genes, and the number of enriched genes following enrichment analysis of gene ontology terms for walleye (*Sander vitreus*) sampled across the Lake Winnipeg system in 2017 and 2018.

Location	Contrast	Regulation	Annotated genes	Unique genes	Enriched genes	
Red River	Both years	Up	43	40	6	
		Down	104	90	50	
	2017 only	Up	89	82	14	
		Down	641	582	451	
	2018 only	Up	248	234	0	
		Down	224	206	126	
	Matheson Island	Both years	Up	0	0	0
			Down	4	3	0
Up vs. R; Down vs. D			1	1	0	
2017 only		Up	27	27	0	
		Down	22	22	0	
		Up vs. R; Down vs. D	3	3	0	
		Down vs. R; Up vs. D	2	2	0	
2018 only		Up	33	32	8	
		Down	35	34	4	
		Up vs. R; Down vs. D	33	26	0	
		Down vs. R; Up vs. D	20	18	5	
Dauphin River		Both years	Up	34	30	4
	Down		9	9	0	
	2017 only	Up	64	62	0	
		Down	25	25	0	
	2018 only	Up	417	373	176	
		Down	331	312	25	

R, Red River; D, Dauphin River

Table S4. Differentially regulated genes in walleye (*Sander vitreus*) sampled from the Red River (R) Matheson Island (M), and Dauphin River (D) in 2017 (17) and 2018 (18). Log₂ fold-change (LogFC), false discovery rate (FDR), annotation source, and uniprot ID, as well as E value, percent ID, and transcript sequence are provided in the .xlsx file.

Table S5. Differentially regulated genes associated with significantly enriched gene ontology (GO) terms (biological processes, BP; molecular functions, MF; cellular components, CC) for walleye (*Sander vitreus*) sampled from the Red River (R) Matheson Island (M), and Dauphin River (D) in 2017 (17) and 2018 (18). The GO category (GO cat.; biological process, BP; molecular function, MF; cellular component, CC), GO representative summary term (GO rep.), GO term description (GO desc.), as well as gene and protein names are presented in the .xlsx file.

Table S6. Differentially regulated genes associated with glycolysis and gluconeogenic pathways, metabolism, mTOR pathway, and hypoxic response in walleye (*Sander vitreus*) sampled from the Red River, Matheson Island, and Dauphin River in 2017 and 2018.

Gene id	Gene	Contrast	D.17vsR.17		D.18vsR.18		M.17vsR.17		M.18vsR.18		D.17vsM.17		D.18vsM.18	
			logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR
<i>Glycolysis/gluconeogenesis</i>														
walleye_DN102551_c0_g1	<i>pck1</i>	R.18 up	-0.92	0.033	-1.84	0.000	-0.17	0.816	-1.03	0.031	-0.75	0.223	-0.81	0.082
walleye_DN113477_c6_g3	<i>aldocb</i>	R.17 down, D.18 up	0.51	0.010	0.67	0.000	0.41	0.048	0.17	0.580	0.10	0.915	0.50	0.019
walleye_DN109278_c2_g1	<i>eno1</i>	R.17 down, D.18 up	0.56	0.014	0.95	0.000	0.54	0.024	0.25	0.447	0.03	0.990	0.70	0.005
walleye_DN113326_c7_g6	<i>gapdh</i>	R.17 down, D.18 up	0.38	0.036	0.36	0.031	0.46	0.014	-0.04	0.925	-0.07	0.944	0.40	0.038
walleye_DN113371_c0_g1	<i>ldha</i>	R.17 down, D.18 up	1.33	0.001	1.09	0.002	1.45	0.000	0.20	0.778	-0.12	0.963	0.89	0.027
walleye_DN113371_c4_g1	<i>ldha</i>	R.17 down, D.18 up	0.93	0.002	0.98	0.000	1.00	0.001	0.23	0.633	-0.07	0.978	0.75	0.018
walleye_DN108570_c4_g3	<i>tpi1b</i>	R.17 down, D.18 up	0.49	0.015	0.66	0.000	0.44	0.037	0.18	0.551	0.05	0.974	0.48	0.026
walleye_DN111971_c3_g8	<i>eno3</i>	D.18 up	0.53	0.040	0.86	0.000	0.49	0.072	0.19	0.663	0.05	0.986	0.67	0.013
walleye_DN105332_c1_g6	<i>fbp1</i>	D.18 up	0.34	0.224	0.57	0.013	0.41	0.132	0.00	0.996	-0.08	0.966	0.58	0.033
walleye_DN111191_c5_g1	<i>hk2</i>	D.18 up	0.33	0.040	0.36	0.014	0.29	0.086	0.01	0.986	0.04	0.975	0.35	0.041
walleye_DN104637_c6_g1	<i>pfkfb1</i>	D.18 up	0.21	0.561	0.85	0.001	0.14	0.767	0.13	0.809	0.08	0.971	0.72	0.018
walleye_DN109466_c13_g1	<i>pkm</i>	D.18 up	0.30	0.156	0.49	0.006	0.55	0.007	-0.12	0.733	-0.25	0.492	0.62	0.004
<i>Metabolism</i>														
walleye_DN104552_c6_g1	<i>mt-nd4</i>	R.both up, M.18 int., D.18 down	-0.43	0.011	-0.85	<0.001	-0.45	0.011	-0.40	0.037	0.02	0.992	-0.45	0.012
walleye_DN109940_c8_g1	<i>mt-nd5</i>	R.both up, M.18 int., D.18 down	-0.66	0.000	-1.17	<0.001	-0.48	0.009	-0.72	0.000	-0.19	0.623	-0.45	0.016
walleye_DN109940_c8_g3	<i>mt-nd5</i>	R.both up, M.18 int., D.18 down	-0.56	0.002	-1.00	<0.001	-0.41	0.027	-0.54	0.008	-0.15	0.769	-0.47	0.016
walleye_DN109940_c8_g7	<i>mt-nd5</i>	R.both up, M.18 int., D.18 down	-0.68	0.000	-1.13	<0.001	-0.50	0.004	-0.63	0.001	-0.19	0.584	-0.50	0.005
walleye_DN109940_c7_g12	<i>mt-nd6</i>	R.both up, M.18 int., D.18 down	-0.65	0.000	-1.10	<0.001	-0.48	0.009	-0.60	0.003	-0.17	0.690	-0.50	0.009
walleye_DN104552_c6_g4	<i>mt-nd3</i>	R.both up	-0.44	0.008	-0.92	<0.001	-0.36	0.037	-0.59	0.002	-0.08	0.922	-0.33	0.066
walleye_DN108955_c4_g10	<i>mt-cyb</i>	R.18 up,	-0.31	0.098	-0.81	<0.001	-0.36	0.052	-0.40	0.047	0.06	0.963	-0.41	0.030

		M.18 int. up, D.18 down												
walleye_DN112225_c12_g2	<i>mt-atp8</i>	R.18 up	-0.18	0.237	-0.46	<0.001	-0.26	0.082	-0.35	0.028	0.08	0.905	-0.11	0.557
walleye_DN108955_c4_g9	<i>mt-cyb</i>	R.18 up	-0.27	0.171	-0.77	<0.001	-0.35	0.071	-0.43	0.036	0.08	0.936	-0.34	0.094
walleye_DN107218_c0_g1	<i>mt-col</i>	R.17 up, D.18 down	-0.49	0.002	-0.68	<0.001	-0.54	0.001	-0.31	0.090	0.05	0.960	-0.36	0.036
walleye_DN107218_c2_g1	<i>mt-col</i>	R.17 up, D.18 down	-0.42	0.011	-0.64	<0.001	-0.51	0.003	-0.28	0.149	0.10	0.881	-0.36	0.042
walleye_DN107248_c63_g4	<i>mt-nd1</i>	R.17 up, D.18 down	-0.67	0.001	-1.09	<0.001	-0.49	0.019	-0.37	0.116	-0.18	0.720	-0.72	0.002
walleye_DN106456_c2_g1	<i>mt-col</i>	R.17 up	-0.43	0.004	-0.61	<0.001	-0.54	0.001	-0.30	0.079	0.11	0.808	-0.31	0.055
walleye_DN110148_c1_g1	<i>mt-col</i>	R.17 up	-0.60	0.002	-0.65	<0.001	-0.63	0.002	-0.31	0.182	0.03	0.986	-0.34	0.109
walleye_DN100513_c0_g1	<i>mterf3</i>	D.18 up	0.25	0.070	0.28	0.021	0.38	0.005	-0.11	0.577	-0.14	0.636	0.40	0.005
walleye_DN102460_c0_g1	<i>atp5h</i>	D.18 down	0.12	0.649	-0.62	0.001	0.07	0.842	-0.20	0.473	0.05	0.971	-0.42	0.040
walleye_DN112225_c12_g1	<i>mt-atp6</i>	D.18 down	-0.43	0.013	-0.84	<0.001	-0.29	0.127	-0.30	0.145	-0.14	0.761	-0.54	0.004
walleye_DN107218_c1_g1	<i>mt-col</i>	D.18 down	-0.43	0.059	-0.61	0.003	-0.47	0.041	-0.12	0.771	0.04	0.984	-0.49	0.041
walleye_DN107248_c63_g5	<i>mt-co2</i>	D.18 down	-0.27	0.225	-0.82	<0.001	-0.17	0.518	-0.27	0.288	-0.09	0.926	-0.55	0.010
walleye_DN112462_c1_g1	<i>mt-co3</i>	D.18 down	-0.52	0.029	-0.84	<0.001	-0.33	0.213	-0.31	0.327	-0.19	0.780	-0.53	0.039
walleye_DN112462_c1_g3	<i>mt-co3</i>	D.18 down	-0.36	0.069	-0.86	<0.001	-0.24	0.274	-0.31	0.187	-0.11	0.880	-0.56	0.006
walleye_DN108955_c3_g4	<i>mt-cyb</i>	D.18 down	-0.27	0.202	-0.84	<0.001	-0.36	0.076	-0.41	0.061	0.09	0.921	-0.43	0.038
walleye_DN104552_c7_g6	<i>mt-nd4</i>	D.18 down	-0.45	0.008	-0.86	<0.001	-0.42	0.017	-0.36	0.063	-0.03	0.984	-0.50	0.006
<i>Hypoxia</i>														
walleye_DN110302_c3_g6	<i>nos1</i>	R.18 down	3.69	0.021	4.90	0.001	3.25	0.051	3.88	0.022	0.44	0.960	1.02	0.552
walleye_DN110766_c1_g5	<i>egln3</i>	R.17 down	1.12	0.047	0.26	0.707	1.19	0.038	-0.11	0.939	-0.07	0.988	0.36	0.641
walleye_DN113735_c3_g1	<i>slc2a1</i>	R.17 down	0.45	0.002	0.41	0.002	0.48	0.002	0.28	0.096	-0.03	0.984	0.13	0.477
walleye_DN113735_c3_g8	<i>slc2a1</i>	R.17 down	0.94	0.005	0.72	0.021	0.76	0.029	0.18	0.784	0.17	0.913	0.54	0.144
walleye_DN100261_c0_g1	<i>hmox</i>	D.18 up	0.24	0.614	0.74	0.027	0.16	0.784	-0.45	0.351	0.08	0.980	1.19	0.003
walleye_DN109309_c3_g1	<i>hyou1</i>	D.18 up	0.48	0.063	0.61	0.007	0.90	0.001	-0.01	0.988	-0.42	0.247	0.63	0.018
walleye_DN113024_c7_g1	<i>hbad</i>	D.18 down	-1.09	0.006	-1.45	0.000	-0.43	0.378	-0.09	0.930	-0.66	0.263	-1.36	0.002
<i>mTOR pathway</i>														
walleye_DN105880_c11_g2	<i>prkaa2</i>	R.18 up	-0.34	0.334	-1.27	0.000	0.13	0.791	-0.73	0.034	-0.48	0.316	-0.54	0.113
walleye_DN105603_c3_g10	<i>seh1l</i>	R.18 down	0.39	0.002	0.56	0.000	0.24	0.073	0.31	0.029	0.15	0.503	0.25	0.061
walleye_DN107366_c3_g3	<i>EIF4A1</i>	R.17 down, D.18 up	0.51	0.001	0.65	0.000	0.40	0.010	0.30	0.079	0.11	0.813	0.35	0.030
walleye_DN111020_c1_g2	<i>EIF4B</i>	R.17 down, D.18 up	0.31	0.023	0.42	0.001	0.38	0.008	0.10	0.656	-0.06	0.932	0.32	0.029
walleye_DN104291_c1_g1	<i>EIF4G1</i>	R.17 down, D.18 up	0.77	0.003	0.91	0.000	0.77	0.005	0.21	0.624	0.00	0.999	0.70	0.012
walleye_DN113534_c6_g1	<i>hras</i>	R.17 down, D.18 up	0.46	0.002	0.53	0.000	0.35	0.022	0.14	0.543	0.11	0.822	0.39	0.012
walleye_DN111314_c0_g4	<i>atf4</i>	R.17 down	0.20	0.037	0.22	0.015	0.33	0.001	0.15	0.196	-0.13	0.444	0.06	0.643
walleye_DN106983_c0_g1	<i>EIF4E</i>	R.17 down	0.42	0.001	0.14	0.275	0.35	0.006	-0.14	0.378	0.07	0.870	0.28	0.031
walleye_DN106499_c5_g4	<i>grb2</i>	R.17 down	0.47	0.000	0.09	0.377	0.32	0.002	0.10	0.460	0.14	0.368	-0.01	0.961

walleye_DN113534_c6_g4	<i>hras</i>	R.17 down	0.28	0.006	0.19	0.043	0.44	0.000	0.11	0.432	-0.16	0.283	0.08	0.568
walleye_DN108382_c6_g1	<i>kras</i>	R.17 down	0.43	0.000	0.25	0.020	0.50	0.000	0.09	0.656	-0.07	0.883	0.17	0.210
walleye_DN109684_c2_g3	<i>lamtor2</i>	R.17 down	0.31	0.006	0.24	0.020	0.25	0.030	0.14	0.364	0.06	0.912	0.10	0.468
walleye_DN105554_c3_g7	<i>mapk1</i>	R.17 down	0.33	0.015	0.31	0.013	0.29	0.039	0.09	0.731	0.04	0.969	0.23	0.131
walleye_DN113739_c6_g6	<i>mapk1</i>	R.17 down	0.51	0.002	0.17	0.319	0.37	0.030	-0.03	0.954	0.14	0.764	0.20	0.306
walleye_DN103901_c0_g1	<i>mtor</i>	R.17 down	0.33	0.012	0.30	0.011	0.32	0.018	0.18	0.290	0.01	0.994	0.13	0.435
walleye_DN105466_c8_g1	<i>pdpk1</i>	R.17 down	0.28	0.006	0.02	0.884	0.36	0.001	0.09	0.593	-0.07	0.824	-0.06	0.672
walleye_DN109353_c2_g8	<i>rragc</i>	R.17 down	0.23	0.017	0.21	0.023	0.26	0.010	0.15	0.234	-0.03	0.967	0.06	0.674
walleye_DN109353_c2_g5	<i>rragc</i>	D.18 up	0.55	0.001	0.56	0.000	0.14	0.526	0.10	0.757	0.40	0.060	0.46	0.009
walleye_DN112490_c3_g1	<i>nprl3</i>	D.18 down	-0.18	0.463	-0.39	0.037	-0.23	0.335	0.18	0.556	0.05	0.974	-0.58	0.008
<i>PI3K/AKT/mTOR pathway</i>														
walleye_DN112562_c7_g4	<i>foxo1a</i>	R.17 down	0.54	0.016	0.29	0.205	0.66	0.005	0.29	0.331	-0.11	0.917	0.00	0.995
walleye_DN103115_c0_g1	<i>acaca</i>	R.17 down, D.18 up	0.68	0.003	1.10	0.000	0.69	0.004	0.37	0.181	-0.01	0.999	0.73	0.004
walleye_DN108540_c0_g1	<i>acly</i>	R.17 down, D.18 up	0.65	0.000	0.83	0.000	0.68	0.000	0.31	0.100	-0.03	0.984	0.52	0.003
walleye_DN108540_c0_g2	<i>acly</i>	R.17 down, D.18 up	0.62	0.000	0.81	0.000	0.77	0.000	0.27	0.212	-0.16	0.717	0.54	0.004
<i>Citric acid cycle</i>														
walleye_DN107782_c2_g6	<i>idh1</i>	R.17 down	1.11	0.001	0.45	0.154	0.94	0.006	0.04	0.971	0.17	0.912	0.42	0.263
walleye_DN107754_c3_g2	<i>ogdh</i>	R.17 down	0.35	0.012	0.15	0.298	0.37	0.011	0.06	0.825	-0.02	0.988	0.09	0.668
walleye_DN107308_c5_g1	<i>sdha</i>	R.17 down	0.35	0.008	-0.06	0.742	0.30	0.029	-0.06	0.814	0.05	0.948	0.01	0.985

R.17, Red River 2017; R.18, Red River 2018; R.both, Red River both years; D.17, Dauphin River 2017; D.18, Dauphin River 2018; M.17, Matheson Island 2017; logFC, Log₂ fold-change; FDR, false discovery rate.

Significant differentially regulated genes are represented as bolded text. logFC values for significantly regulated genes are coloured to reflect genes differentially regulated in Red River (teal) and Dauphin River (blue) fish. In some cases, multiple contrasts may have been significant.

Table S7. Differentially regulated genes associated with ion regulation in walleye (*Sander vitreus*) sampled from the Red River, Matheson Island, and Dauphin River in 2017 and 2018.

Gene id	Gene	Contrast	D.17vsR.17		D.18vsR.18		M.17vsR.17		M.18vsR.18		D.17vsM.17		D.18vsM.18	
			logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR	logFC	FDR
walleye_DN108377_c4_g1	<i>aqp3</i>	M.17 up	0.04	0.987	1.56	0.288	3.62	0.009	1.32	0.540	-3.58	0.035	0.24	0.927
walleye_DN111678_c3_g2	<i>aqp8</i>	D.18 down	-1.48	0.098	-2.90	0.001	-0.57	0.641	-0.75	0.570	-0.92	0.618	-2.15	0.031
walleye_DN102608_c0_g1	<i>aqpa</i>	D.18 down	-1.14	0.013	-1.99	0.000	-0.59	0.263	-0.73	0.188	-0.55	0.535	-1.26	0.010
walleye_DN113771_c9_g2	<i>atp1a1</i>	R.18 up	-0.08	0.856	-1.16	0.000	0.17	0.692	-0.76	0.016	-0.25	0.725	-0.40	0.198
walleye_DN104577_c1_g1	<i>atp1a1</i>	R.17 up	-0.59	0.009	-0.84	0.000	-0.52	0.027	-0.44	0.094	-0.07	0.963	-0.41	0.101
walleye_DN104577_c1_g2	<i>atp1a1</i>	D.18 down	-0.22	0.651	-1.66	0.000	0.11	0.866	-0.47	0.310	-0.33	0.705	-1.19	0.002
walleye_DN113771_c14_g1	<i>atp1a1</i>	D.18 down	-0.11	0.843	-1.50	0.000	0.18	0.748	-0.49	0.292	-0.30	0.760	-1.01	0.008
walleye_DN113771_c14_g3	<i>atp1a1</i>	D.18 down	-0.09	0.890	-1.52	0.000	0.17	0.778	-0.43	0.394	-0.26	0.832	-1.09	0.007
walleye_DN106483_c6_g4	<i>atp1a3</i>	R.17 down	0.80	0.002	0.69	0.004	0.58	0.031	0.21	0.617	0.22	0.754	0.48	0.089
walleye_DN113771_c11_g1	<i>atp1a3</i>	D.18 up	0.23	0.140	0.54	0.000	0.23	0.141	0.08	0.787	-0.01	0.997	0.47	0.003
walleye_DN113771_c11_g5	<i>atp1a3</i>	D.18 up	0.26	0.292	0.77	0.000	0.36	0.133	0.23	0.459	-0.10	0.937	0.54	0.022
walleye_DN113859_c9_g1	<i>atp1a3</i>	D.18 up	0.20	0.190	0.41	0.002	0.24	0.109	-0.02	0.960	-0.04	0.965	0.43	0.005
walleye_DN113488_c5_g3	<i>atp2a2</i>	D.both down	-0.42	0.016	-0.39	0.017	0.03	0.921	0.00	0.996	-0.45	0.041	-0.39	0.039
walleye_DN106163_c0_g1	<i>atp2b4</i>	R.17 down	0.72	0.000	0.52	0.001	0.58	0.001	0.23	0.286	0.14	0.748	0.30	0.102
walleye_DN103456_c0_g1	<i>atp2b4</i>	M.18 up	2.63	0.267	-4.18	0.144	1.35	0.669	5.29	0.030	1.29	0.849	-9.47	0.003
walleye_DN111653_c1_g1	<i>atp6v0a2</i>	R.17 down	0.25	0.044	0.19	0.110	0.37	0.004	-0.03	0.933	-0.12	0.681	0.22	0.110
walleye_DN107940_c2_g1	<i>atp6v0e1</i>	R.17 down	0.33	0.002	0.10	0.386	0.23	0.035	0.01	0.970	0.10	0.704	0.09	0.542
walleye_DN106269_c5_g1	<i>atp6v1a</i>	R.17 down, D.18 up	1.22	0.000	1.24	0.000	0.87	0.002	0.43	0.184	0.36	0.436	0.82	0.005
walleye_DN102031_c0_g1	<i>atp6v1b2</i>	R.17 down, D.18 up	0.43	0.002	0.49	0.000	0.34	0.020	0.13	0.514	0.10	0.827	0.36	0.016
walleye_DN108788_c7_g1	<i>atp6v1c1a</i>	R.17 down, D.18 up	0.47	0.001	0.51	0.000	0.39	0.005	0.08	0.766	0.07	0.897	0.43	0.004
walleye_DN104855_c4_g3	<i>atp6v1c1a</i>	D.18 up	0.49	0.014	0.56	0.003	0.19	0.465	0.07	0.873	0.30	0.349	0.49	0.024
walleye_DN110109_c4_g3	<i>atp6v1e1</i>	R.17 down, D.18 up	0.68	0.000	0.76	0.000	0.48	0.017	0.16	0.588	0.21	0.618	0.60	0.004
walleye_DN106672_c2_g2	<i>atp6v1h</i>	D.17 down	-6.37	0.008	0.51	0.846	0.10	0.980	0.40	0.932	-6.46	0.035	0.10	0.982
walleye_DN104183_c4_g7	<i>camk1</i>	R.both down	0.56	0.000	0.65	0.000	0.64	0.000	0.58	0.001	-0.08	0.920	0.07	0.781
walleye_DN106317_c1_g1	<i>camk1d</i>	D.18 up	1.39	0.011	2.38	0.000	0.64	0.328	0.60	0.439	0.75	0.414	1.79	0.003
walleye_DN105627_c1_g1	<i>camk2g</i>	R.17 down	0.38	0.004	0.18	0.172	0.31	0.020	0.05	0.865	0.06	0.923	0.13	0.435
walleye_DN104069_c3_g1	<i>cdc42</i>	R.both down	0.41	0.000	0.34	0.000	0.43	0.000	0.33	0.005	-0.02	0.987	0.01	0.959
walleye_DN104069_c3_g2	<i>cdc42</i>	R.both down	0.40	0.002	0.25	0.034	0.55	0.000	0.29	0.043	-0.15	0.550	-0.04	0.861
walleye_DN107058_c0_g1	<i>clcn5</i>	R.17 down	0.50	0.015	0.17	0.436	0.51	0.015	0.07	0.876	-0.02	0.994	0.10	0.750
walleye_DN111682_c2_g1	<i>cldn1</i>	D.18 down	0.03	0.947	-0.49	0.041	0.52	0.054	0.26	0.476	-0.48	0.175	-0.75	0.006
walleye_DN111682_c2_g6	<i>cldn1</i>	D.18 down	0.30	0.432	-0.60	0.039	0.79	0.014	0.16	0.785	-0.49	0.301	-0.76	0.023
walleye_DN110492_c2_g9	<i>cldn22</i>	R.both up	-1.14	0.013	-0.96	0.023	-1.43	0.003	-1.11	0.032	0.28	0.869	0.15	0.855
walleye_DN100552_c0_g1	<i>cldn3</i>	R.17 down	4.05	0.046	1.05	0.633	4.38	0.037	4.64	0.044	-0.33	0.984	-3.58	0.091
walleye_DN106664_c5_g3	<i>cldn4</i>	D.18 down	-0.06	0.901	-0.63	0.017	0.39	0.221	0.09	0.894	-0.45	0.298	-0.72	0.019

walleye_DN83550_c0_g1	<i>cldn5</i>	R.both up	-0.69	0.010	-1.07	0.000	-1.13	0.000	-1.05	0.001	0.44	0.285	-0.02	0.977
walleye_DN110590_c3_g1	<i>cldn8</i>	D.18 down	-0.51	0.174	-1.46	0.000	0.05	0.939	-0.62	0.132	-0.56	0.291	-0.85	0.023
walleye_DN103849_c4_g3	<i>kcjnl</i>	R.18 up	-0.64	0.188	-1.23	0.003	-0.31	0.623	-1.15	0.023	-0.33	0.815	-0.08	0.934
walleye_DN104249_c1_g4	<i>kcjnl16</i>	D.18 down	0.00	0.997	-1.06	0.002	0.26	0.616	-0.04	0.967	-0.26	0.827	-1.02	0.009
walleye_DN111684_c8_g2	<i>nr3c2</i>	R.17 down	0.65	0.011	0.74	0.002	0.67	0.011	0.51	0.083	-0.02	0.994	0.23	0.475
walleye_DN104639_c0_g1	<i>rhcgl</i>	R.18 up, M.18 Int, D.18 down	-0.54	0.123	-2.07	0.000	-0.09	0.882	-0.77	0.041	-0.45	0.436	-1.31	0.001
walleye_DN112974_c2_g2	<i>slc12a3</i>	D.18 up	0.35	0.688	1.15	0.047	-0.49	0.563	-0.45	0.674	0.84	0.437	1.60	0.018
walleye_DN106771_c6_g3	<i>slc12a8</i>	D.18 up	0.59	0.219	1.74	0.000	-0.19	0.798	0.20	0.825	0.78	0.220	1.54	0.002
walleye_DN105800_c0_g5	<i>slc4a1</i>	R.17 up	-1.83	0.012	-1.21	0.080	-1.53	0.043	-0.82	0.414	-0.30	0.941	-0.39	0.731
walleye_DN111084_c5_g1	<i>slc9a1</i>	R.18 up	-0.90	0.002	-1.71	0.000	-0.44	0.177	-1.10	0.001	-0.46	0.312	-0.61	0.057
walleye_DN105658_c3_g2	<i>slc9a3</i>	R.18 up, M18 Int, D.18 down	-0.16	0.764	-1.92	0.000	-0.07	0.918	-0.81	0.037	-0.09	0.974	-1.11	0.004
walleye_DN108637_c1_g1	<i>tjp3</i>	R.17 down	0.38	0.012	0.42	0.003	0.45	0.004	0.28	0.119	-0.07	0.929	0.14	0.485

R.17, Red River 2017; R.18, Red River 2018; R.both, Red River both years; D.17, Dauphin River 2017; D.18, Dauphin River 2018; M.17, Matheson Island 2017; logFC, Log₂ fold-change; FDR, false discovery rate

Significant differentially regulated genes are represented as bolded text. logFC values for significantly regulated genes are coloured to reflect genes differentially regulated in Red River (teal), Matheson Island (light blue), and Dauphin River (dark blue) fish. In some cases, multiple contrasts may have been significant.

Table S8. Results for linear models examining the consistency of candidate gene mRNA levels measured by qPCR of walleye sampled in the Lake Winnipeg system (Red River, Matheson Island, Dauphin River) in 2017 and 2018 that were or were not part of the RNA-seq study.

Gene	Factor^a	Sum squares	df	F-value	p-value
<i>actn4</i>	Year	6136.00	1	8.46	0.005
	RNAseq	3954.00	1	5.45	0.022
	Total length	78.00	1	0.11	0.744
	Mass	41.00	1	0.06	0.814
<i>actr2</i>	Location	3.77	2	7.06	0.001
	Year	1.48	1	5.57	0.020
	Total length	0.10	1	0.37	0.546
<i>akr1a1b</i>	Year	6.00	1	19.46	< 0.001
	RNAseq	1.12	1	3.65	0.059
<i>ap1s1</i>	Location	1.20	2	2.52	0.086
	Year	2.08	1	8.75	0.004
	Total length	0.11	1	0.46	0.498
	Location × Year	0.95	2	2.01	0.141
<i>arf6</i>	Location	3.818	2	12.06	< 0.001
	Year	0.07	1	0.44	0.510
	Total length	0.02	1	0.10	0.755
	Location × Year	1.76	2	5.55	0.005
<i>arhgdia</i>	Year	0.79	1	2.63	0.108
	RNAseq	0.67	1	2.23	0.138
	Total length	0.06	1	0.21	0.652
	Mass	0.38	1	1.24	0.268
<i>cdc42</i>	Location	0.60	2	1.87	0.160
	Year	1.35	1	8.40	0.005
	RNAseq	0.17	1	1.05	0.309
<i>cnnm4</i>	Location	0.32	2	0.79	0.457
	Year	3.43	1	16.73	< 0.001
	RNAseq	0.50	1	2.45	0.121
	Total length	0.21	1	1.04	0.310
	Location × Year	1.14	2	2.78	0.067
<i>cot1l</i>	Location	16.90	2	42.46	< 0.001
	Year	0.34	1	1.72	0.193
	Total length	0.08	1	0.38	0.538
	Mass	0.00	1	0.02	0.879
<i>dgka</i>	Location	1.99	2	3.56	0.033
	Year	8.29	1	29.61	< 0.001
	RNAseq	0.87	1	3.12	0.081
	Total length	0.59	1	2.10	0.151
<i>dnajc9</i>	Location	1.39	2	3.59	0.031
	Year	3.56	1	18.36	< 0.001
	RNAseq	0.38	1	1.93	0.168

	Total length	0.00	1	0.01	0.931
<i>eipr1</i>	Location	0.99	2	1.98	0.144
	Year	3.38	1	13.57	< 0.001
	RNAseq	0.70	1	2.81	0.097
	Total length	0.25	1	1.02	0.316
	Location × Year	0.82	2	1.65	0.197
<i>fam49b</i>	Location	3.67	2	9.68	< 0.001
	Year	3.95	1	20.82	< 0.001
	RNAseq	0.23	1	1.21	0.275
	Mass	0.00	1	0.02	0.887
	Location × Year	0.91	2	2.40	0.097
<i>gpx1</i>	Location	7.21	2	12.69	< 0.001
	Year	7.59	1	26.72	< 0.001
	Mass	0.18	1	0.62	0.435
	Location × Year	1.41	2	2.48	0.089
<i>mcu</i>	Location	1.56	2	2.97	0.056
	Year	4.34	1	16.54	< 0.001
	RNAseq	1.01	1	3.84	0.053
	Total length	0.61	1	2.32	0.132
	Mass	0.34	1	1.29	0.259
<i>pfn2</i>	Location	2.63	2	5.69	0.005
	Mass	0.21	1	0.92	0.339
<i>rpa2</i>	Location	9.96	2	15.68	< 0.001
	RNAseq	0.05	1	0.17	0.679
<i>slc25a24</i>	Location	0.95	2	1.41	0.248
	Year	5.40	1	16.14	< 0.001
<i>tnfaip8l2b</i>	Year	3.28	1	16.02	< 0.001
	RNAseq	0.20	1	0.98	0.324
<i>ube2j1</i>	Location	3.78	2	7.81	< 0.001
	Year	0.29	1	1.19	0.279
	Mass	0.15	1	0.60	0.440
	Location × Year	4.33	2	8.94	< 0.001

^a Akaike Information Criterion (AIC) was used to determine which fixed factors (location, year, RNAseq vs. non-RNAseq, total length, mass, location × year) were included in the linear model. Significant fixed factors are represented as bolded text. See Table S2 for gene abbreviations.

Table S9. Results for the linear models examining the relative mRNA levels measured by qPCR of 20 candidate genes for walleye (*Sander vitreus*) sampled from the Red River, Riverton, Matheson Island, and Dauphin River in 2017 and 2018.

Gene	Factor^a	Sum squares	df	F-value	p-value
<i>actn4</i>	Location	1.32	3	1.45	0.232
	Year	5.42	1	17.87	< 0.001
<i>actr2</i>	Location	23909.00	3	5.18	0.002
	Year	8163.00	1	5.31	0.023
	Mass	320.00	1	0.21	0.649
<i>akr1a1b</i>	Location	1.95	3	2.31	0.079
	Year	1.57	1	5.58	0.020
	Mass	0.04	1	0.15	0.698
	Location × Year	2.27	3	2.69	0.049
<i>ap1s1</i>	Location	1.35	3	1.95	0.124
	Year	2.46	1	10.68	0.001
	Total length	0.12	1	0.53	0.468
<i>arf6</i>	Location	3.86	3	8.37	< 0.001
	Year	0.007	1	0.47	0.496
	Total length	0.02	1	0.14	0.710
	Location × Year	1.93	3	4.18	0.007
<i>arhgdia</i>	Year	1.41	1	4.85	0.029
	Mass	0.35	1	1.21	0.273
<i>cdc42</i>	Location	1.02	3	1.90	0.132
	Year	1.44	1	8.06	0.005
	Mass	0.04	1	0.23	0.630
<i>cnm4</i>	Location	2.14	3	3.53	0.017
	Year	0.08	1	0.38	0.537
	Total length	0.30	1	1.50	0.222
	Location × Year	1.87	3	3.08	0.030
<i>cot11</i>	Location	18.78	3	28.65	< 0.001
	Year	1.24	1	5.66	0.019
	Total length	0.12	1	0.54	0.464
	Mass	0.01	1	0.02	0.875
<i>dgka</i>	Location	2.15	3	2.73	0.046
	Year	9.10	1	34.77	< 0.001
	Total length	0.06	1	0.24	0.627
<i>dnajc9</i>	Location	1.98	3	2.80	0.042
	Year	3.79	1	16.11	< 0.001
	Mass	0.27	1	1.15	0.286
<i>eipr1</i>	Location	1.66	3	2.03	0.112
	Year	3.83	1	14.07	< 0.001
	Mass	0.172	1	0.63	0.427
<i>fam49b</i>	Location	5.46	3	9.87	< 0.001

	Year	4.65	1	25.24	< 0.001
	Mass	1.05	1	5.67	0.018
	Location × Year	0.86	3	1.55	0.205
<i>gpx1</i>	Location	9.23	3	9.86	< 0.001
	Year	10.28	1	32.91	< 0.001
	Location × Year	1.58	3	1.69	0.173
<i>mcu</i>	Location	1.58	3	2.05	0.110
	Year	5.22	1	20.32	< 0.001
	Total length	0.008	1	0.03	0.861
<i>pfn2</i>	Location	2.86	3	4.25	0.007
	Year	0.08	1	0.37	0.542
<i>rpa2</i>	Location	9.12	3	8.97	< 0.001
	Total length	0.02	1	0.05	0.817
<i>slc25a24</i>	Location	1.91	3	2.13	0.100
	Year	7.70	1	25.88	< 0.001
	Mass	0.34	1	1.13	0.290
<i>tnfaip8l2b</i>	Location	0.84	3	1.42	0.240
	Year	2.71	1	13.77	< 0.001
	Mass	0.32	1	1.60	0.207
<i>ube2j1</i>	Location	3.88	3	5.73	0.001
	Year	0.30	1	1.32	0.253
	Mass	0.33	1	1.47	0.227
	Location × Year	4.57	3	6.74	< 0.001

^a Akaike Information Criterion (AIC) was used to determine which fixed factors (location, year, total length, mass, location × year) were included in the linear model.

Significant fixed factors are represented as bolded text. See Table S2 for gene abbreviations.

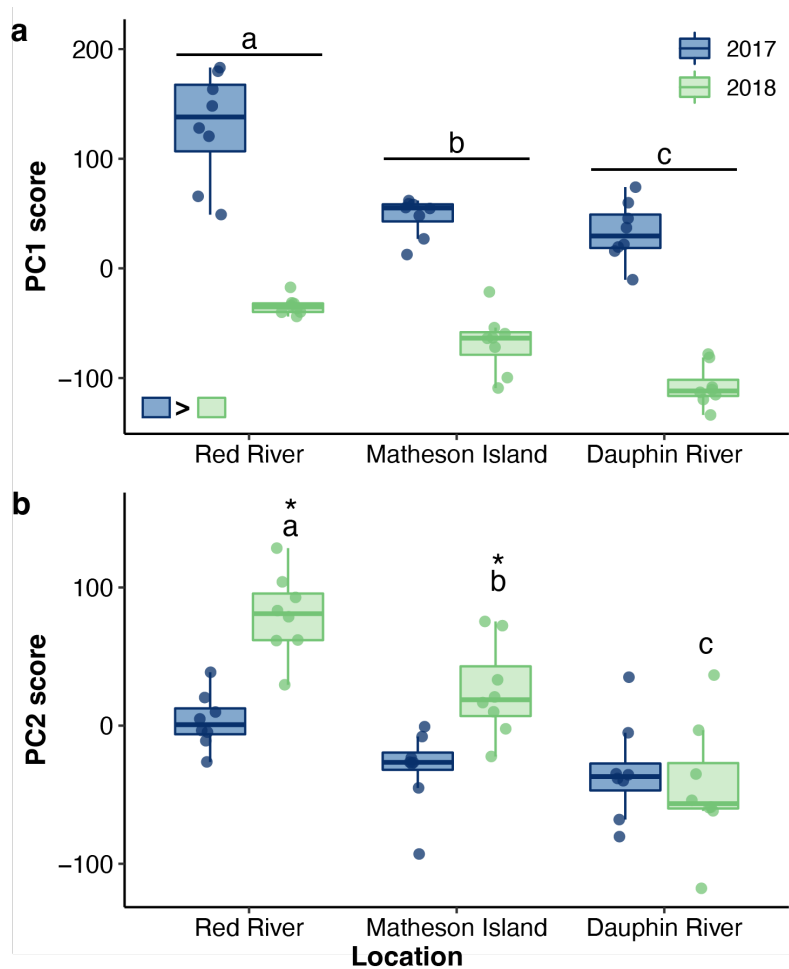


Figure S1. Principal component (PC) scores for PC1 (a) and PC2 (b) for whole-transcriptomic analysis of walleye (*Sander vitreus*) sampled in the Lake Winnipeg system in 2017 and 2018 ($n = 8$). Walleye were sampled from the Red River, Matheson Island, and Dauphin River, representing sites in the south basin, channel, and north basin, respectively. Locations that do not share a letter are significantly different from one another. An asterisk represents a significant effect of year within a location, while the inset represents an overall significant effect of year across locations (two-way ANOVA; see text for details). Horizontal bars in the boxplot represent the median response value and 75%, 50%, and 25% quartiles. Whiskers represent ± 1.5 times the interquartile range, and each dot represents an individual response value.

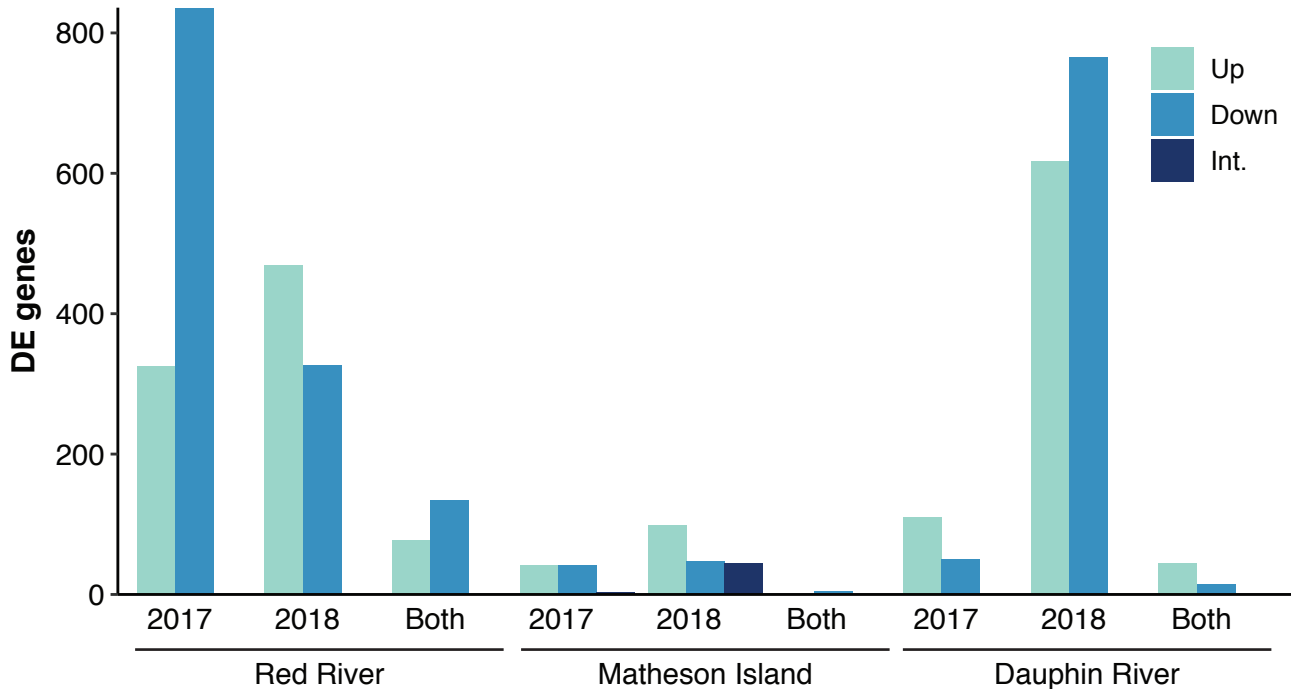


Figure S2. Number genes that were differentially expressed (DE) in walleye (*Sander vitreus*) from the Lake Winnipeg system in 2017 or 2018 only, or consistently across both years (Both). Walleye were sampled in the Red River, Matheson Island, and Dauphin River, representing sites in the south basin, channel, and north basin, respectively. Site comparisons represent differences of one site compared to the other two sites (i.e., Red River compared to both Matheson Island and Dauphin River; Matheson Island compared to both Red River and Dauphin River; Dauphin River compared to both Red River and Matheson Island). Genes are expressed in terms of their direction of regulation (up or down). Intermediate (int.) represents regulation in opposite directions relative to the two other sampling sites (e.g., for Matheson Island, up-regulated compared to Red River, down-regulated compared to Dauphin River).

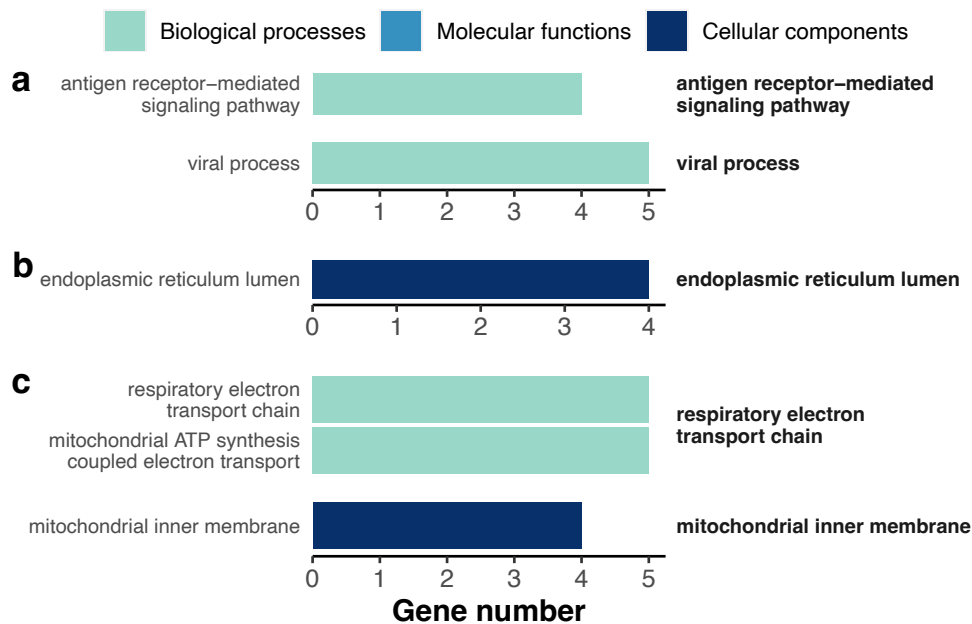


Figure S3. Summary of the enriched gene ontology (GO) terms of genes down-regulated in 2018 only for walleye (*Sander vitreus*; $n = 8$) sampled from Matheson Island compared to Red River and Dauphin River. Genes were considered differentially regulated at a false discovery rate < 0.05 . Only GO terms from the functional analysis with an adjusted $p < 0.05$ with more than four transcripts were considered as significantly enriched. Significant GO terms were summarized using REVIGO to reduce redundancy and grouped according to similarity (right labels).

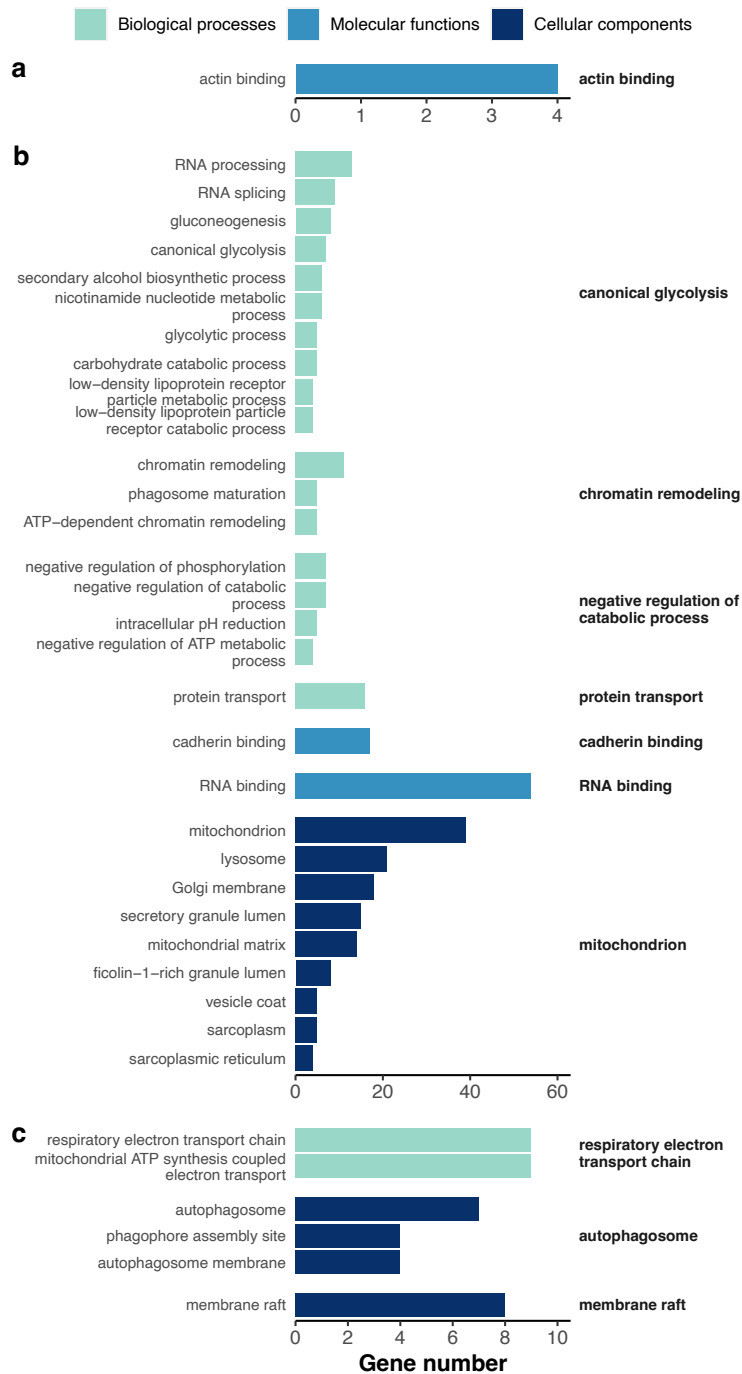


Figure S4. Summary of the enriched gene ontology (GO) terms of genes (a) up-regulated in both 2017 and 2018 as well as (b) up- and (c) down-regulated in 2018 only for walleye (*Sander vitreus*; $n = 8$) sampled from Dauphin River compared to Red River and Matheson Island. Genes were considered differentially regulated at a false discovery rate < 0.05 . Only GO terms from the functional analysis with an adjusted $p < 0.05$ with more than four transcripts were considered as significantly enriched. Significant GO terms were summarized using REVIGO to reduce redundancy and grouped according to similarity (right labels).

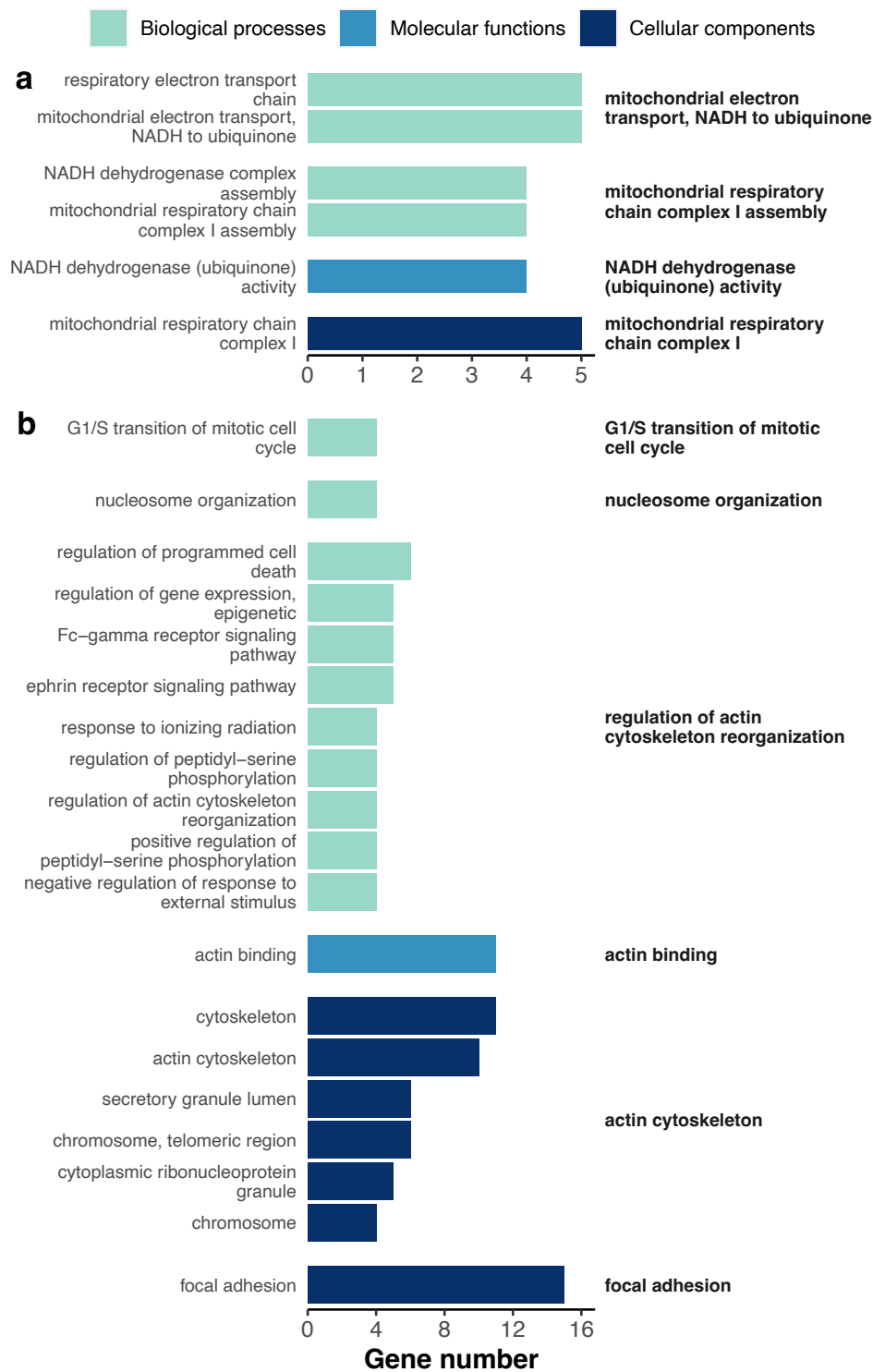


Figure S5. Summary of the enriched gene ontology (GO) terms of genes (a) up- and (b) down-regulated in both 2017 and 2018 for walleye (*Sander vitreus*; $n = 8$) sampled from the Red River compared to Matheson Island and Dauphin River. Genes were considered differentially regulated at a false discovery rate < 0.05 . Only GO terms from the functional analysis with an adjusted $p < 0.05$ with at least four transcripts were considered as significantly enriched. Significant GO terms were summarized using REVIGO to reduce redundancy and grouped according to similarity (right labels).

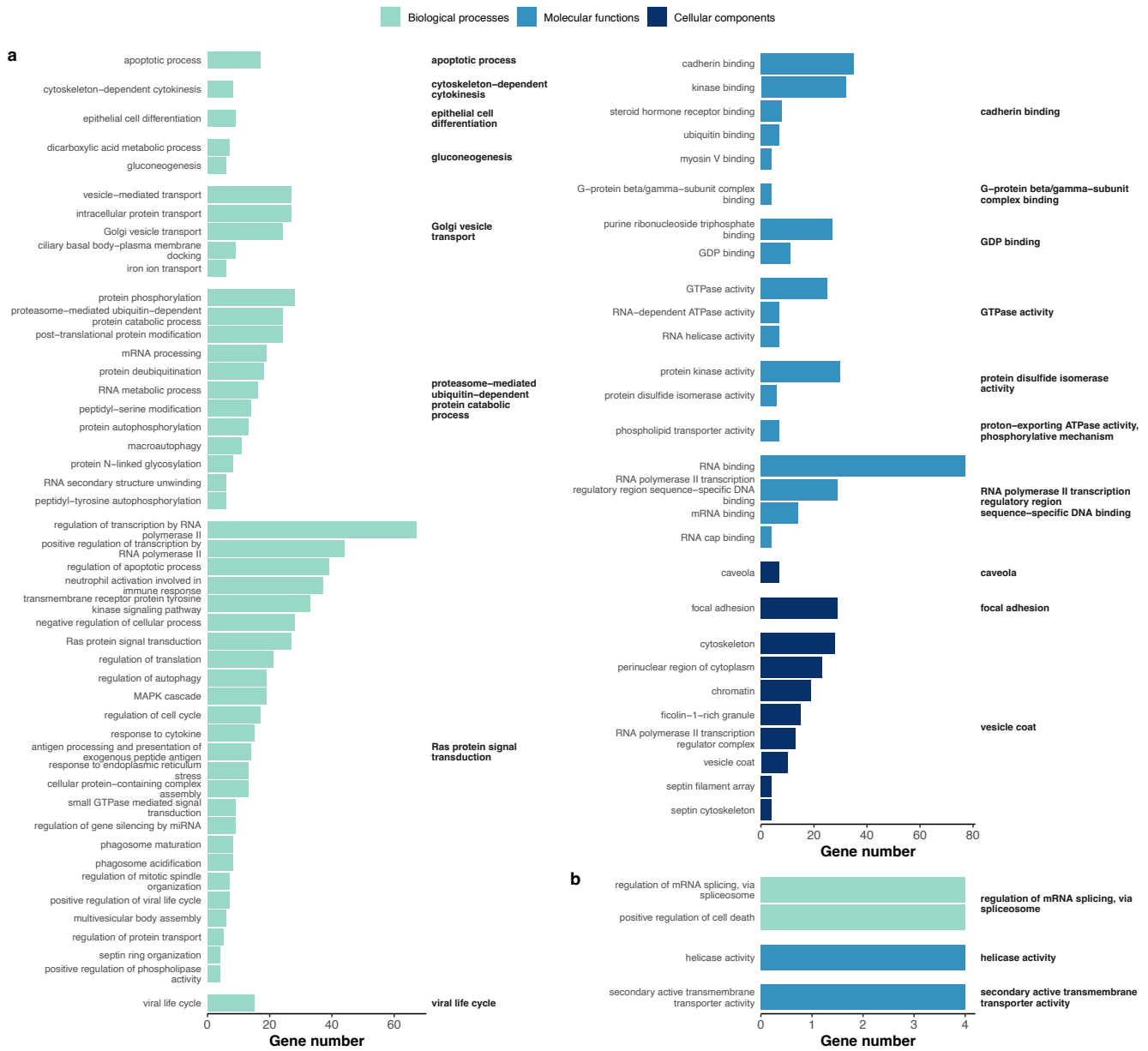


Figure S6. Summary of the enriched gene ontology (GO) terms of genes (a) down- and (b) up-regulated in 2017 only for walleye (*Sander vitreus*; $n = 8$) sampled from the Red River compared to Matheson Island and Dauphin River. Genes were considered differentially regulated at a false discovery rate < 0.05 . Only GO terms from the functional analysis with an adjusted $p < 0.05$ with more than four transcripts were considered as significantly enriched. Significant GO terms were summarized using REVIGO to reduce redundancy and grouped according to similarity (right labels).

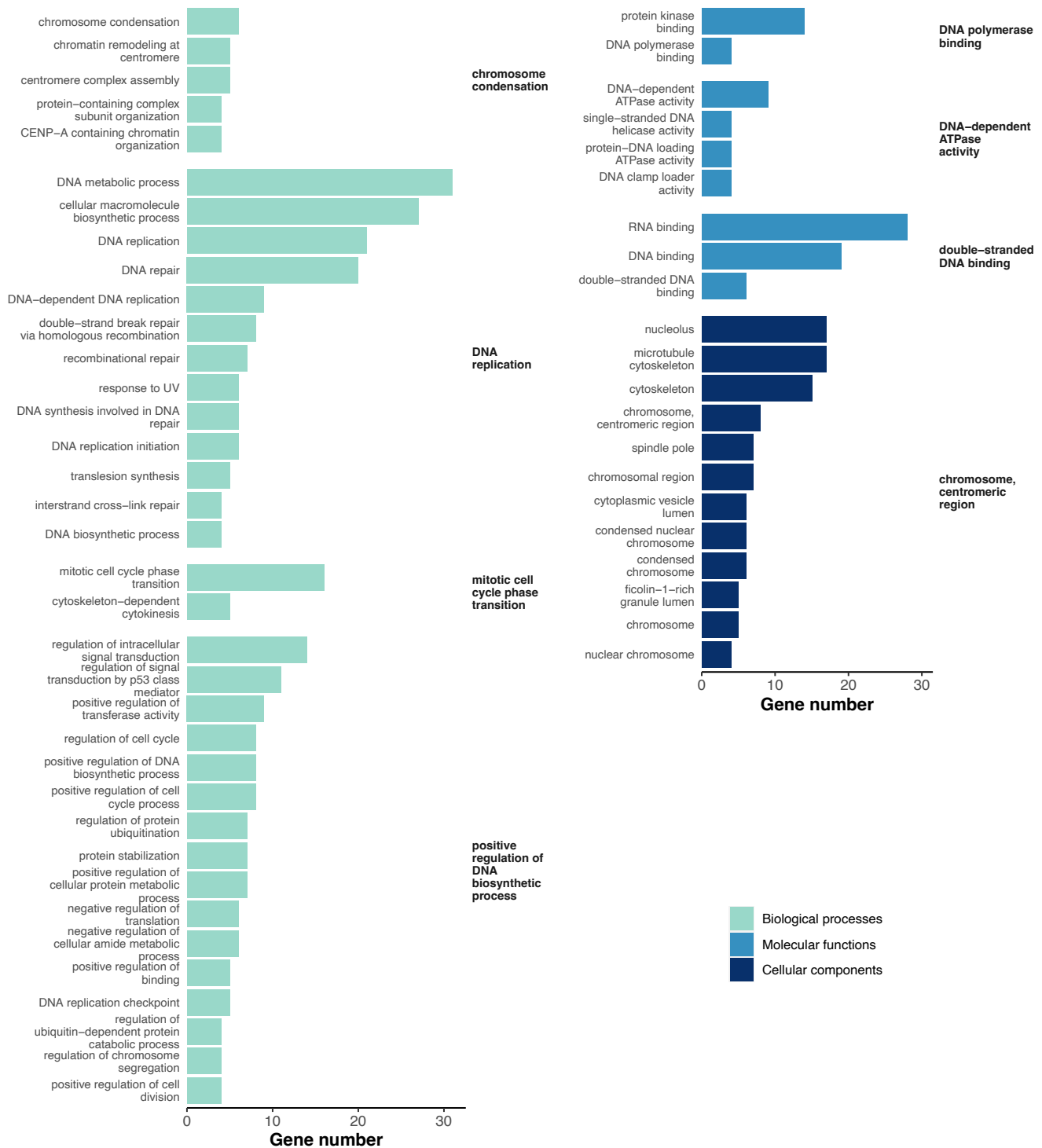


Figure S7. Summary of the enriched gene ontology (GO) terms of genes down-regulated in 2018 only for walleye (*Sander vitreus*; $n = 8$) sampled from the Red River compared to Matheson Island and Dauphin River. Genes were considered differentially regulated at a false discovery rate < 0.05 . Only GO terms from the functional analysis with an adjusted $p < 0.05$ with more than four transcripts were considered as significantly enriched. Significant GO terms were summarized using REVIGO to reduce redundancy and grouped according to similarity (right labels).

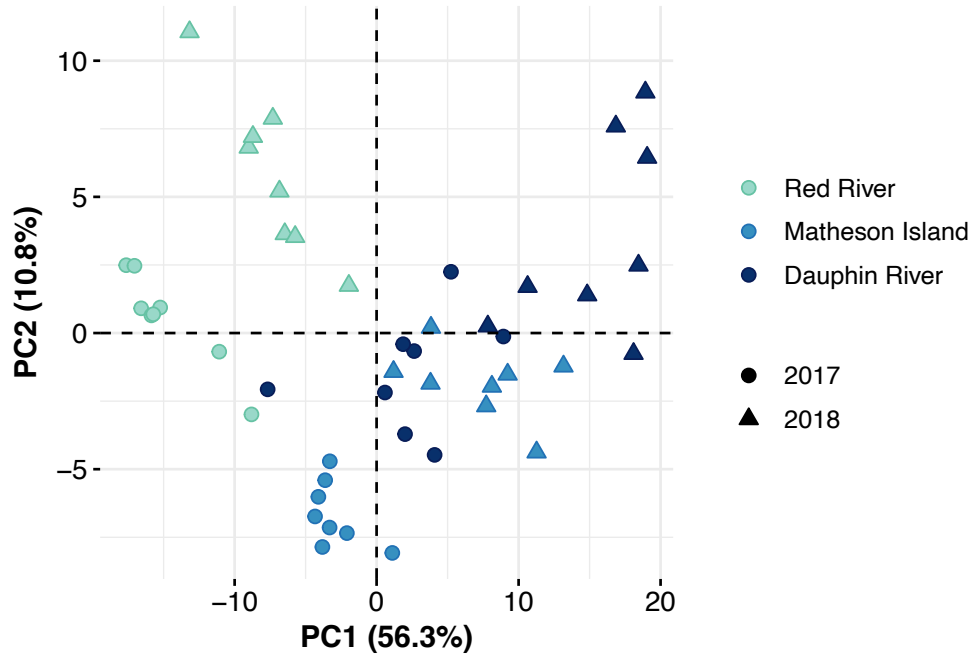


Figure S8. Principal components analysis (PCA) of the 195 annotated genes that were differentially regulated across both 2017 and 2018 in walleye (*Sander vitreus*) sampled in the Lake Winnipeg system. Walleye were sampled from the Red River, Matheson Island, and Dauphin River in 2017 and 2018 ($n = 8$ per site and year), and genes that were differentially regulated ($FDR < 0.05$) at one site compared to the other two sites were identified. The variance explained by each PC is indicated in brackets.

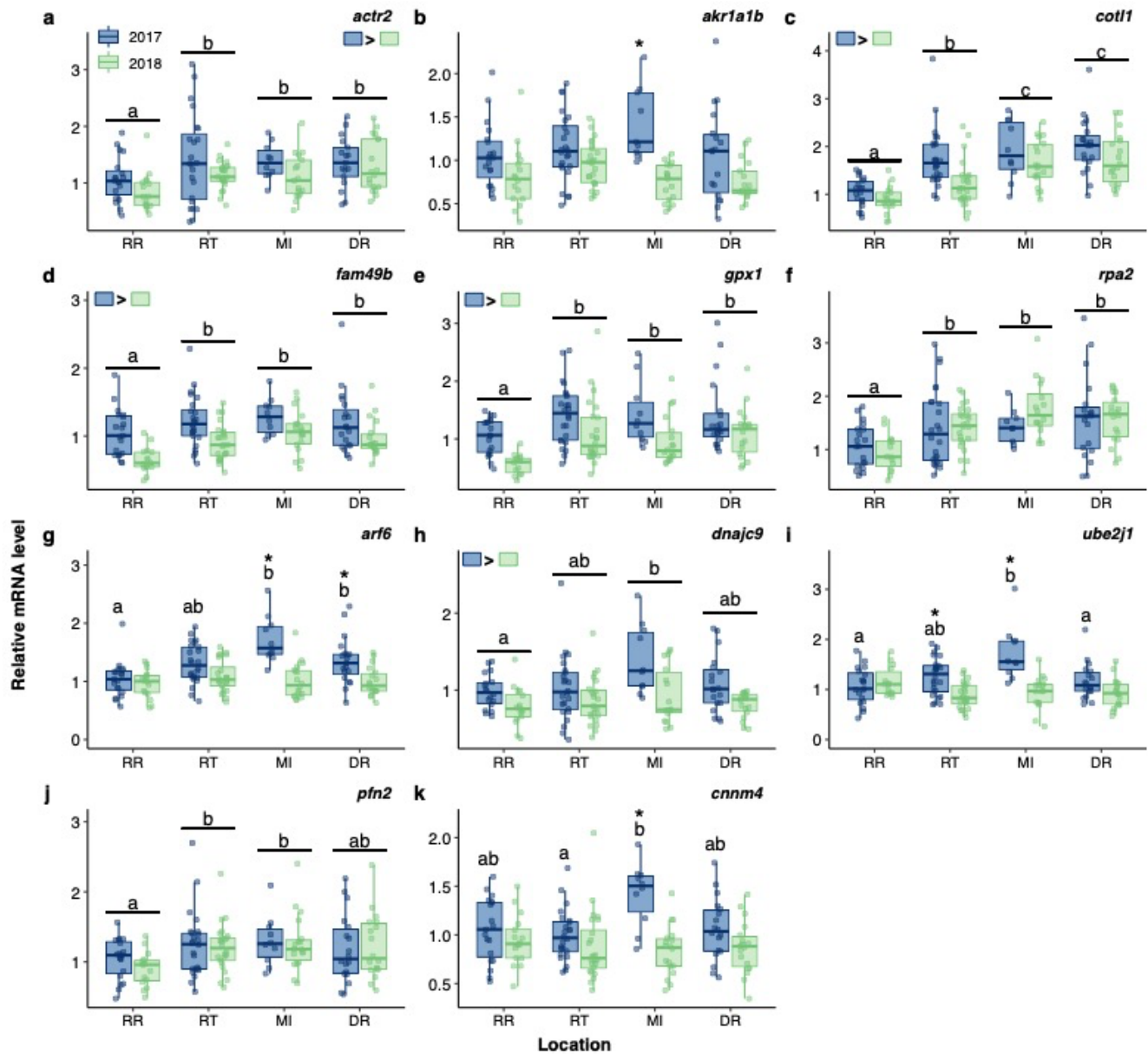


Figure S9. Relative mRNA levels for candidate genes of walleye (*Sander vitreus*) from the lake Winnipeg system. Walleye were sampled from the Red River (RR; $n = 14-19$), Riverton (RT; $n = 23-24$), Matheson Island (MI; $n = 9-18$), and Dauphin River (DR; $n = 17-19$) in 2017 and 2018. Locations that do not share a letter are significantly different from one another. An asterisk represents a significant effect of year within a location, while the inset represents an overall significant effect of year across locations (see Table S9). Horizontal bars in the boxplot represent the median response value and 75%, 50%, and 25% quartiles. Whiskers represent ± 1.5 times the interquartile range, and each dot represents an individual response value. See Table S2 for gene abbreviations.

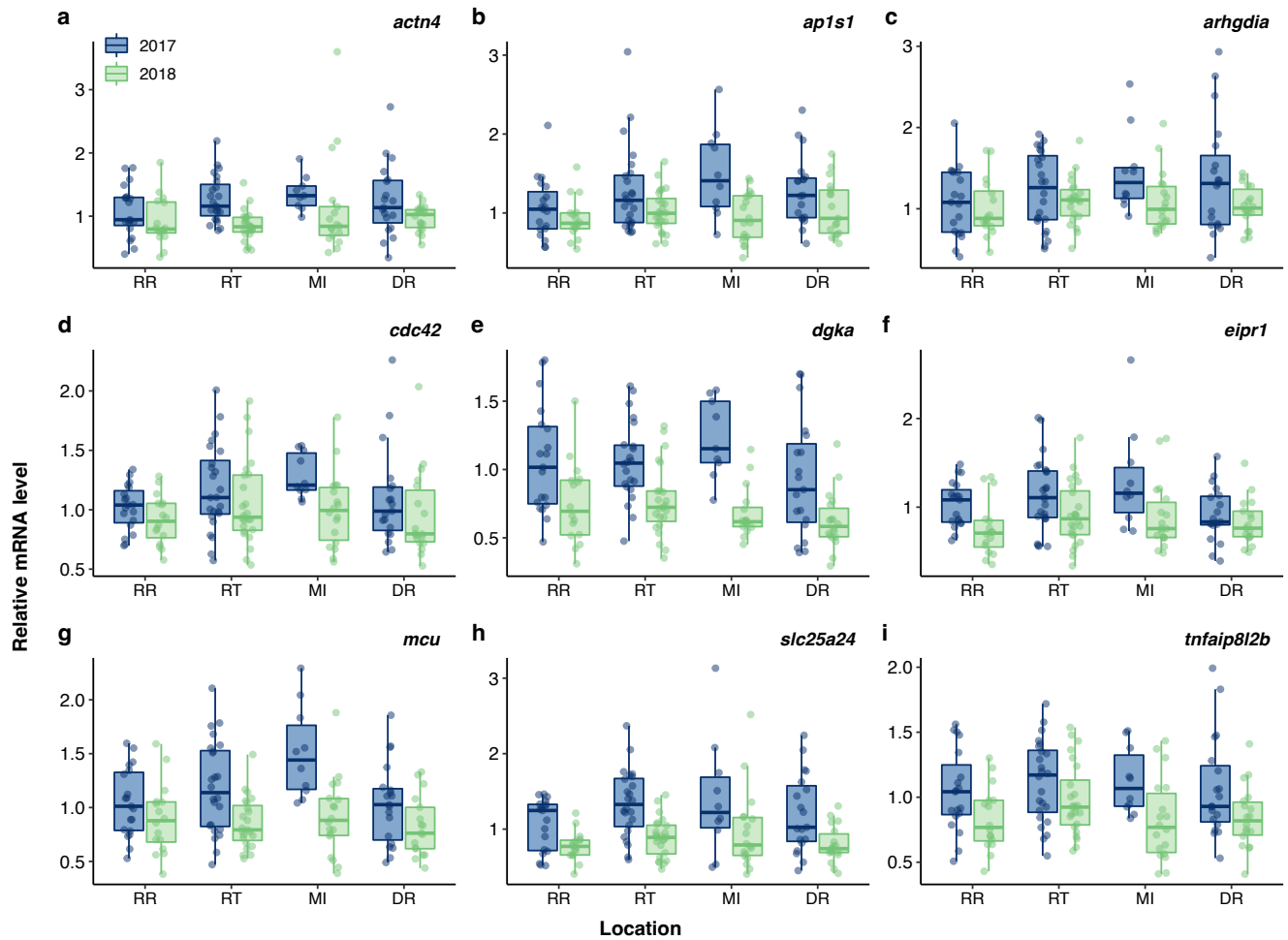


Figure S10. Relative mRNA levels for candidate genes of walleye (*Sander vitreus*) from the lake Winnipeg system. Walleye were sampled from the Red River (RR; $n = 16-19$), Riverton (RT; $n = 22-24$), Matheson Island (MI; $n = 9-18$), and Dauphin River (DR; $n = 17-19$) in 2017 and 2018. For each gene, there was a significant effect of year but not location (see Table S9). Horizontal bars in the boxplot represent the median response value and 75%, 50%, and 25% quartiles. Whiskers represent ± 1.5 times the interquartile range, and each dot represents an individual response value. See Table S2 for gene abbreviations.